

SUPPLEMENTARY DATA

Supplementary Table S1. Site directed mutation of signal peptide of preproinsulin

Designation	Sequence
Wild type PPI SP sequence	MALWMRLLPLLALLALWGPDPAAA
9L mutant	MALWMRLL <u>LLLLLL</u> LWGPDPAAA
12L mutant	MALWMRLLLLL <u>LLL</u> LWGPDPAAA
15L mutant	MALWMRLLLLLLL <u>L</u> LWGPDPAAA
9L/12L mutant	MALWMRLL <u>LLL</u> LLLWGPDPAAA
9L/15L mutant	MALWMRLL <u>LLLLL</u> LWGPDPAAA
12L/15L mutant	MALWMRLLLLL <u>LL</u> LWGPDPAAA
9L/12L/15L mutant	MALWMRLL <u>LLL</u> LLWGPDPAAA

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Supplementary Table S2. Details of subjects studied for pancreatic expression of antigen processing enzymes

Case ID	Group	Source	Donor Status	Age (years)	Duration
485 88	No diabetes control	UK Pancreas Biobank	Autopsy	2	
315 89	No diabetes control	UK Pancreas Biobank	Autopsy	9	
65 71	No diabetes control	UK Pancreas Biobank	Autopsy	40	
8579	No diabetes control	UK Pancreas Biobank	Autopsy	7	
540 91	No diabetes control	UK Pancreas Biobank	Autopsy	11	
PAN8	No diabetes control	UK Pancreas Biobank	Autopsy	19	
E560	Type 1 diabetes	UK Type 1 diabetes Biobank	Organ Donor	42	1.5y
Sc115	Type 1 diabetes	UK Type 1 diabetes Biobank	Autopsy	1	0 'Recent'
E124B	Type 1 diabetes	UK Type 1 diabetes Biobank	Autopsy	17	0 'Recent'
E375	Type 1 diabetes	UK Type 1 diabetes Biobank	Autopsy	11	unknown
11746	Type 1 diabetes	UK Type 1 diabetes Biobank	Autopsy	6	1 week
11713	Type 1 diabetes	UK Type 1 diabetes Biobank	Autopsy	3	3 months

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Supplementary Table S3. Details of the staining protocol for each antibody employed in the current study. Where multiple antigens were stained on the same section, antibodies were applied sequentially (staining order: anti-TAP1 (overnight at 4°C) followed by anti-HLA-ABC (1h at room temperature) or anti-glucagon (1h at room temperature) then anti-insulin plus DAPI (1h at room temperature)).

Primary Antibody	Manufacturer and clone	Antigen Retrieval	Antibody Dilution	Incubation time with primary antibody	Secondary Detection System
HLA-ABC	Abcam C#ab70328 Mouse monoclonal EMR8-5	10mM citrate pH6.0	1/1000	1h at RT	Immunofluorescence staining using anti-mouse IgG (H+L) Alexa Fluor™-conjugated secondary antibodies (1/400 for 1h)
Insulin	DakoC#A0564 Guinea-pig polyclonal	10mM citrate pH6.0	1/700	1h at RT	Immunofluorescence staining using anti-guinea-pig IgG (H+L) Alexa Fluor™ -conjugated secondary antibodies (1/400 for 1h)
Glucagon	AbcamC#ab92517 Rabbitmonoclonal EP3070	10mM citrate pH6.0	1/4000	1h at RT	Immunofluorescence staining using anti-rabbit IgG (H+L) Alexa Fluor™-conjugated secondary antibodies (1/400 for 1h)
TAP1	Protein Tech C#11114-1-AP Rabbit polyclonal	10mM citrate pH6.0	1/200	o/n 4°C	Immunofluorescence staining using anti-rabbit IgG (H+L) Alexa Fluor™ -conjugated secondary antibodies (1/400 for 1h)
ERAP	R & D Systems C#AF2334 Goat polyclonal	10mM Tris, 1mM EDTA pH9.0	1/200	o/n 4°C	Dako REAL™ Envision™ Detection System with anti-goat IgG HRP-conjugated secondary antibodies (1/800 for 1h).

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Supplementary Table S4. Oligonucleotides used for CRISPR-mediated gene knockout.

	Forward	Reverse
Exon 2	GAAGCGGGCGGCATCCCGGCGTTTT	GCCGGGATGCCGCCCCTTCCGGTG
Exon 3	GCAGGAGGTTGATGTACTCCGTTTT	GGAGTACATCAACCTCCTGCCGGTG
Exon 10	GCAGCCTACATCTTCGGCCTGTTTT	AGGCCGAAGATGTAGGCTGCCGGTG
Exon 11	CAGGACAGGAAAACCGATGCGTTTT	GCATCGGTTTTCTGTCCTGCGGTG

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Supplementary Table S5. Eluted epitopes are not selectively derived from SP containing source protein nor their signal peptide region.

	HLA-A*02:01	HLA-A*11:01	HLA-A*24:02	HLA-B*18:01	HLA-B*38:01	HLA-B*39:06	All HLA	Human proteome
Source proteins	458	667	190	510	378	259	2462	20226
Number of source proteins containing a signal peptide	26	29	17	25	35	21	153	3560
Frequency of signal peptide containing source proteins	5.68%	4.35%	8.95%	4.90%	9.26%	8.11%	6.21%	17.6%
Number of proteins with signal peptide-derived epitope	12	2	4	5	4	13	40	
Frequency of identified SP epitopes from SP-Protein	46.15%	6.90%	23.53%	20.00%	11.43%	61.90%	26.14%	

Source Proteins are those represented by at least one epitope with Mascot Score >40 within our HLA class I elution data (1). Uniprot (2) (access via <http://www.uniprot.org/>) retrieve/ID mapping function was used to identify signal peptide containing source proteins. SP, signal peptide.

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Supplementary Table S6. Putative PPI epitopes identified in silico binding prediction algorithms.

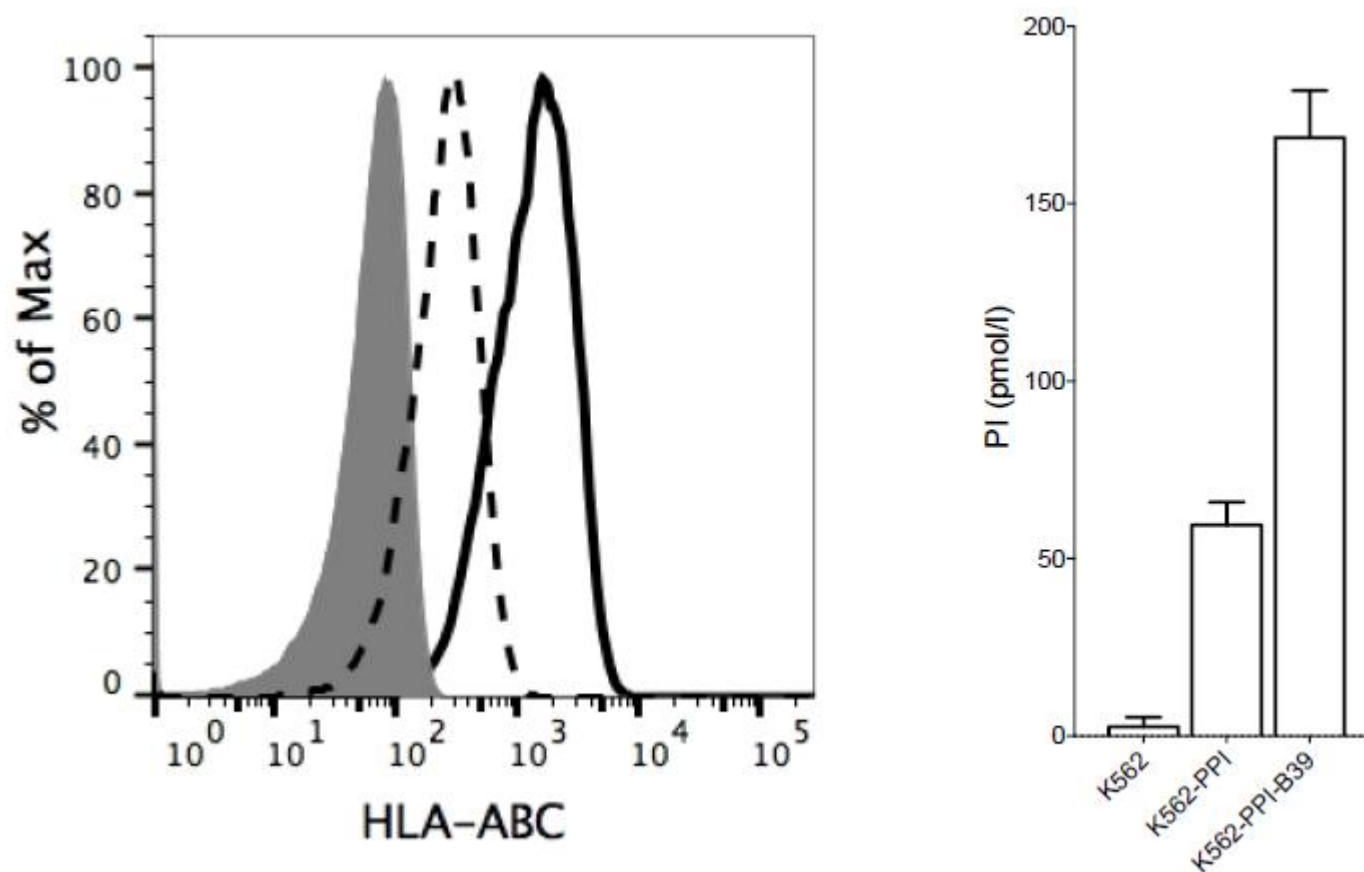
Binding prediction were performed by SYFPEITHI (3) (access via : www.syfpeithi.de) to identify putative PPI epitopes to compare eluted epitopes to those identified in silico. Epitopes derived from the PPI signal peptide are highlighted in bold and those identified by elution in green.

A2	9-mer	Score	A11	9-mer	Score	A24	9-mer	Score	B18	8-mer	Score	B38	9-mer	Score	
1	6	RLLPLLALL	31	38	ALYLVCGER	18	6	RLLPLLALL	15	34	HLVEALYL	15	33	SHLVEALYL	21
2	2	ALWMRLPL	28	81	ALEGSQKR	18	17	WGPDPAAAF	13	36	VEALYLCV	14	5	MRLPLLAL	18
3	34	HLVEALYL	27	35	LVEALYLCV	16	39	LYLVCGERG	13	56	REAEQLQV	14	6	RLLPLLALL	14
4	60	DLQVGQVEL	25	75	GSLQPLALE	16	97	TSICSLYQL	13	61	LQVGQVEL	13	3	LWMRLPLL	13
5	15	ALWGPDPA	22	76	SLQPLALEG	16	3	LWMRLPLL	12	32	GSHLVEAL	12	2	ALWMRLPL	12
6	8	LPLLALLAL	20	32	GSHLVEALY	15	5	MRLPLLAL	12	42	VCGERGFF	12	8	LPLLALLAL	12
7	3	LWMRLPLL	19	80	LALEGSQK	15	8	LPLLALLAL	12	58	AEDLQVGQ	12	31	CGSHLVEAL	12
8	5	MRLPLLAL	19	62	QVGQVELGG	14	22	AAAFVNQHL	12	3	LWMRLPL	11	60	DLQVGQVEL	12
9	81	ALEGSQKR	19	84	GSLQKRGI	14	31	CGSHLVEAL	12	6	RLLPLLAL	11	72	PGAGSLQPL	12
10	7	LPLLALLA	18	85	SLQKRGI	14	33	SHLVEALYL	12	9	PLLALLAL	11	74	AGSLQPLAL	12
11	76	SLQPLALEG	18	96	CTSICSLYQ	14	49	FYTPKTRRE	12	33	SHLVEALY	11	97	TSICSLYQL	12
12	22	AAAFVNQHL	17	2	ALWMRLPL	13	78	QPLALEGSL	12	41	LVCGERGF	11	28	QHLGCSHLV	11
13	69	GGGPGAGSL	17	4	WMRLPLLA	13	2	ALWMRLPL	11	44	GERGFFYT	11	69	GGGPGAGSL	11
14	97	TSICSLYQL	17	7	LPLLALLA	13	53	KTRREAEDL	11	66	VELGGGPG	11	78	QPLALEGSL	11
15	9	PLLALLALW	16	47	GFFYTPKTR	13	69	GGGPGAGSL	11	75	GSLQPLAL	11	94	QCCTSICSL	11
16	10	LLALLALWG	16	10	LLALLALWG	12	74	AGSLQPLAL	11	82	LEGSQKR	11	22	AAAFVNQHL	10
17	31	CGSHLVEAL	16	29	HLCGSHLVE	12	94	QCCTSICSL	11	92	VEQCCTSI	11	91	IVEQCCTSI	10
18	58	AEDLQVGQV	16	60	DLQVGQVEL	12	27	NQHLGCSHL	10	96	CTSICSLY	11	17	WGPDPAAAF	9
19	94	QCCTSICSL	16	98	SICSLYQLE	12	40	YLVCGERGF	10	1	MALWMRL	10	27	NQHLGCSHL	9
20	12	ALLALWGPD	15	12	ALLALWGPD	11	41	LVCGERGF	10	18	GPDPAAAF	10	41	LVCGERGF	9
21	13	LLALWGPD	15	41	LVCGERGF	11	60	DLQVGQVEL	10	43	CGERGFFY	10	53	KTRREAEDL	9
22	18	GPDPAAAFV	15	45	ERGFYTPK	11	72	PGAGSLQPL	10	54	TRREAEDL	10	83	EGSLQKRGI	9
23	28	QHLGCSHLV	15	50	YTPKTRREA	11	83	EGSLQKRGI	10	101	SLYQLENY	10	40	YLVCGERGF	8
24	33	SHLVEALYL	15	53	KTRREAEDL	11	91	IVEQCCTSI	10	4	WMRLPLL	9	55	RREAEDLQV	8
25	38	ALYLVCGER	15	97	TSICSLYQL	11	102	LYQLENYCN	10	7	LPLLALL	9	54	TRREAEDLQ	7

A2	10-mer	Score	A11	10-mer	Score	A24	10-mer	Score	B18	9-mer	Score	B38	10-mer	Score	
1	2	ALWMRLPLL	28	79	PLALEGSQK	21	39	LYLVCGERGF	22	100	CSLYQLENY	15	5	MRLPLLALL	19
2	7	LPLLALLAL	28	75	GSLQPLALEG	20	5	MRLPLLALL	14	60	DLQVGQVEL	14	59	EDLQVGQVEL	14
3	4	WMRLPLLAL	24	84	GSLQKRGI	19	26	VNQHLCGSHL	13	31	CGSHLVEAL	13	7	LPLLALLAL	13
4	29	HLCGSHLVEA	24	32	GSHLVEALYL	18	1	MALWMRLPL	12	44	GERGFFYTP	13	73	GAGSLQPLAL	13
5	15	ALWGPDPA	22	6	RLLPLLALLA	15	2	ALWMRLPLL	12	56	REAEQLQVG	13	1	MALWMRLPL	12
6	1	MALWMRLPL	21	35	LVEALYLVCG	15	7	LPLLALLAL	12	8	LPLLALLAL	12	2	ALWMRLPLL	12
7	90	GIVEQCCTSI	21	85	SLQKRGI	15	30	LCGSHLVEAL	12	36	VEALYLVCG	12	4	WMRLPLLAL	12
8	85	SLQKRGI	20	98	SICSLYQLE	15	59	EDLQVGQVEL	12	58	AEDLQVGQV	12	32	GSHLVEALYL	12
9	6	RLLPLLALLA	19	44	GERGFFYTPK	14	77	LQPLALEGSL	12	74	AGSLQPLAL	12	33	SHLVEALYL	12
10	10	LLALLALWGP	19	9	PLLALLALWG	13	90	GIVEQCCTSI	12	78	QPLALEGSL	12	68	LGGGPGAGSL	12
11	33	SHLVEALYL	19	97	TSICSLYQLE	13	16	LWGPDPAAF	11	82	LEGSQKR	12	93	EQCCTSICSL	12
12	76	SLQPLALEGS	19	29	HLCGSHLVEA	12	21	PAAAFVNQHL	11	3	LWMRLPLL	11	26	VNQHLCGSHL	11
13	96	CTSICSLYQL	19	47	GFFYTPKTRR	12	49	FYTPKTRREA	11	5	MRLPLLAL	11	28	QHLGCSHLVE	11
14	5	MRLPLLALL	18	53	KTRREAEDLQ	12	52	PKTRREAEDL	11	6	RLLPLLALL	11	30	LCGSHLVEAL	11
15	13	LLALWGPDPA	18	2	ALWMRLPLL	11	73	GAGSLQPLAL	11	32	GSHLVEALY	11	52	PKTRREAEDL	11
16	30	LCGSHLVEAL	17	12	ALLALWGPD	11	93	EQCCTSICSL	11	42	VCGERGFFY	11	71	GPAGSLQPL	11
17	73	GAGSLQPLAL	17	15	ALWGPDPA	11	40	YLVCGERGF	10	95	CCTSICSLY	11	96	CTSICSLYQL	11
18	68	LGGGPGAGSL	16	34	HLVEALYLCV	11	68	LGGGPGAGSL	10	9	PLLALLALW	10	16	LWGPDPAAF	10
19	80	LALEGSQKR	16	41	LVCGERGFFY	11	71	GPAGSLQPL	10	27	NQHLGCSHL	10	21	PAAAFVNQHL	10
20	12	ALLALWGPD	15	100	CSLYQLENYC	11	82	LEGSQKRGI	10	40	YLVCGERGF	10	77	LQPLALEGSL	10
21	57	EADLQVGQV	15	4	WMRLPLLAL	10	96	CTSICSLYQL	10	41	LVCGERGF	10	40	YLVCGERGF	9
22	71	GPAGSLQPL	15	7	LPLLALLAL	10	4	WMRLPLLAL	9	66	VELGGGPGA	10	54	TRREAEDLQV	9
23	14	LALWGPDPA	14	25	FVNQHLCGSH	10	32	GSHLVEALYL	9	2	ALWMRLPL	9	39	LYLVCGERGF	8
24	77	LQPLALEGSL	14	37	EALYLVCGER	10	24	AFVNQHLCGS	6	17	WGPDPAAAF	9	45	ERGFYTPKT	8
25	101	SLYQLENYCN	14	46	RGFFYTPKTR	10	42	VCGERGFFY	6	53	KTRREAEDL	9	55	RREAEDLQVG	8

SUPPLEMENTARY DATA

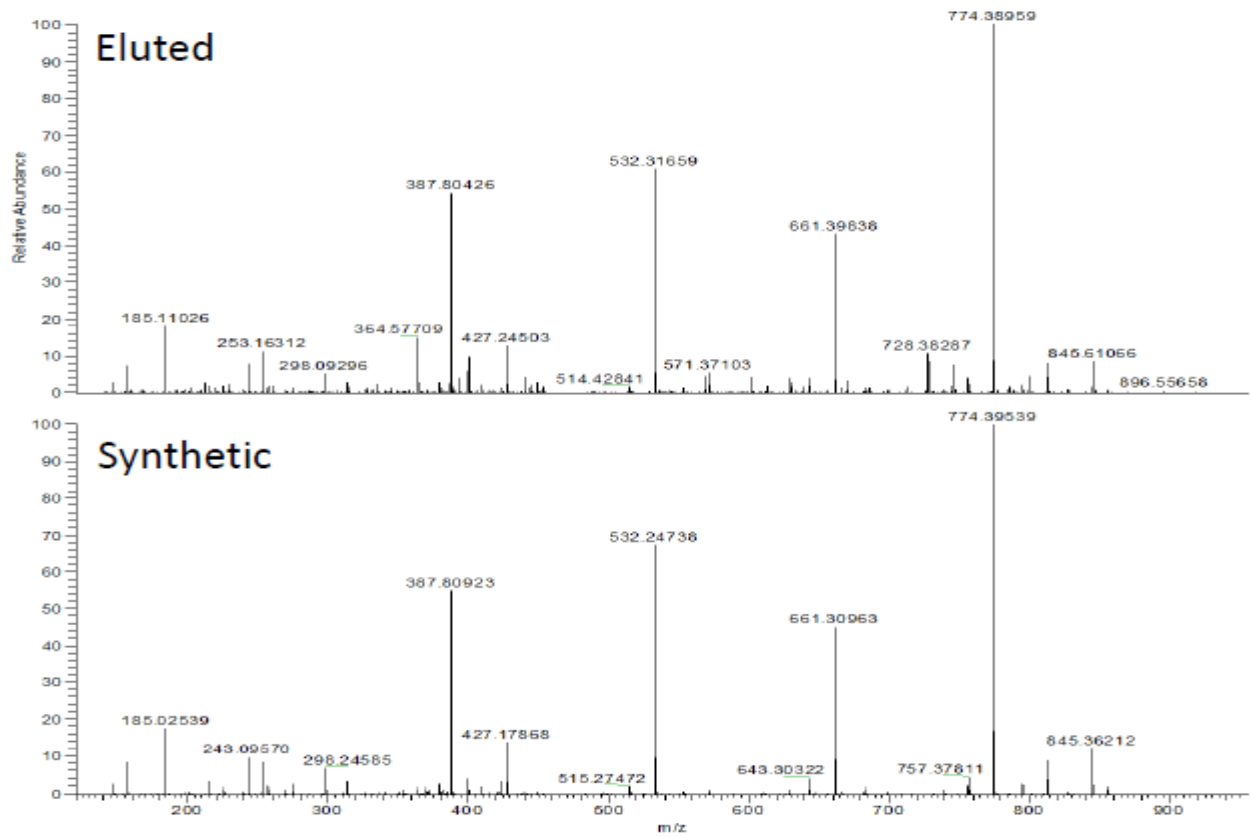
Supplementary Figure S1. Representative Expression of HLA class I and secretion of proinsulin. Expression of HLA class I and secretion of proinsulin by the generated K562-PPI (dashed line) and K562-PPI-HLA (solid line) cell lines (isotype control grey shaded). Single cell cloning was performed to select for the K562-PPI-HLA cell line with best combination of HLA class I expression (left) and Proinsulin expression (right). Data is representative, with B*3906 shown, and similar data obtained for A*11:01, B*1801 and B*38:01.



SUPPLEMENTARY DATA

Supplementary Figure S2. PPI epitope discovery Tandem mass spectrometry analysis of collision-induced dissociation revealing the tandem mass spectrum of a PPI peptide (A) HLA-A*1101 Epitope PPI₈₀₋₈₈ (LALEGLSQQK), (B) HLA-B*3801 Epitope PPI₅₋₁₄ (MRLPLALL) and (C) HLA-B*3801 Epitope PPI₃₃₋₄₁ (SHLVEALYL) . The correct identity of the peptide was proven by tandem mass spectrometry of the synthetic compound. The table lists the amino acid sequence of the peptide with the expected b- and y-fragment ions (fragment ions extending from the amino- and carboxyl terminus respectively). Observed fragment ions are underlined.

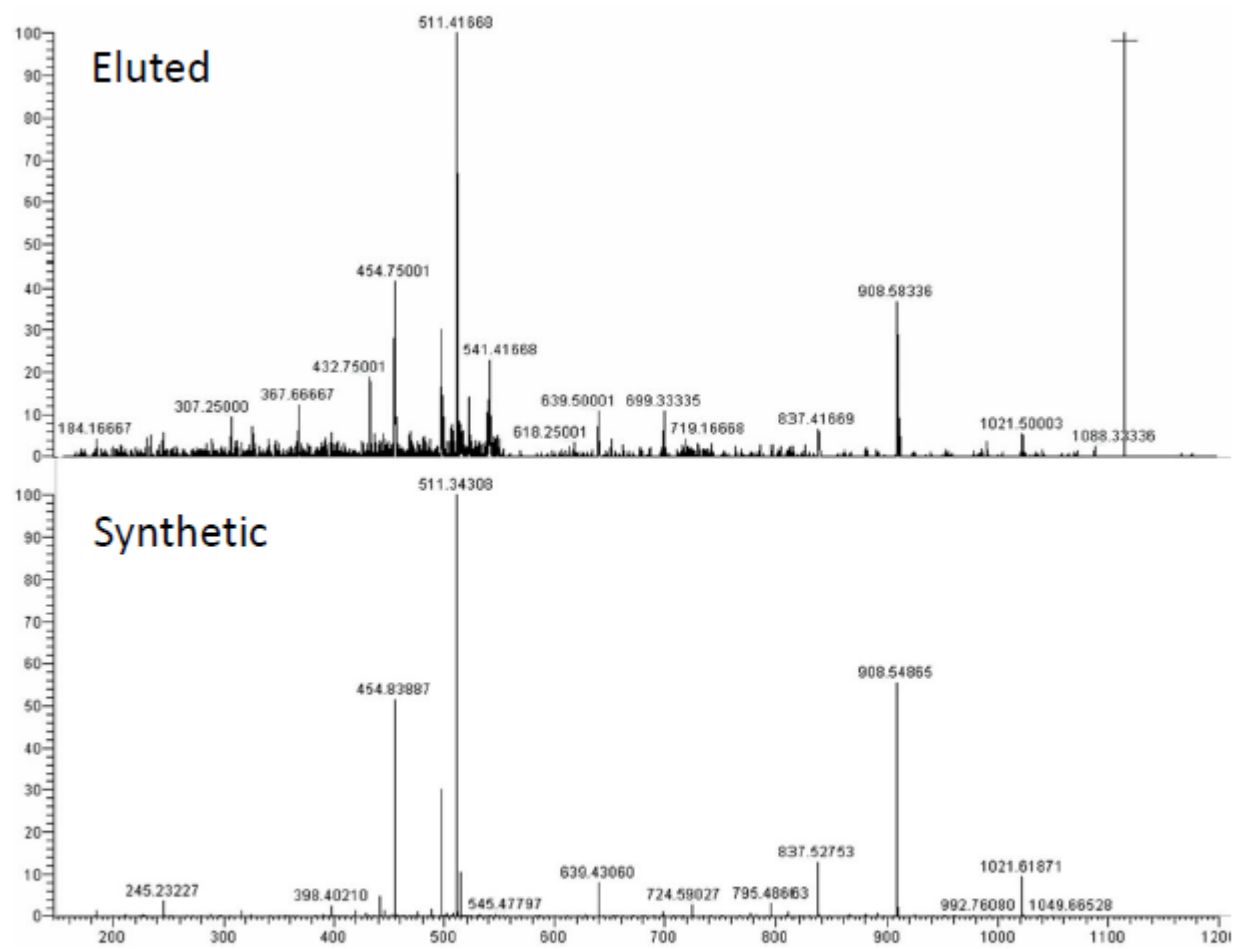
Supplementary Figure 2 A) HLA-A*1101 Epitope PPI₈₀₋₈₈



b _n ions	114.1	<u>185.1</u>	<u>298.2</u>	<u>427.3</u>	484.3	<u>571.3</u>	<u>684.4</u>	<u>812.5</u>	958.6
Peptide	L	A	L	E	G	S	L	Q	K
y _n ions	958.6	<u>845.5</u>	<u>774.4</u>	<u>661.4</u>	<u>523.3</u>	475.3	<u>388.3</u>	<u>275.2</u>	<u>147.1</u>

SUPPLEMENTARY DATA

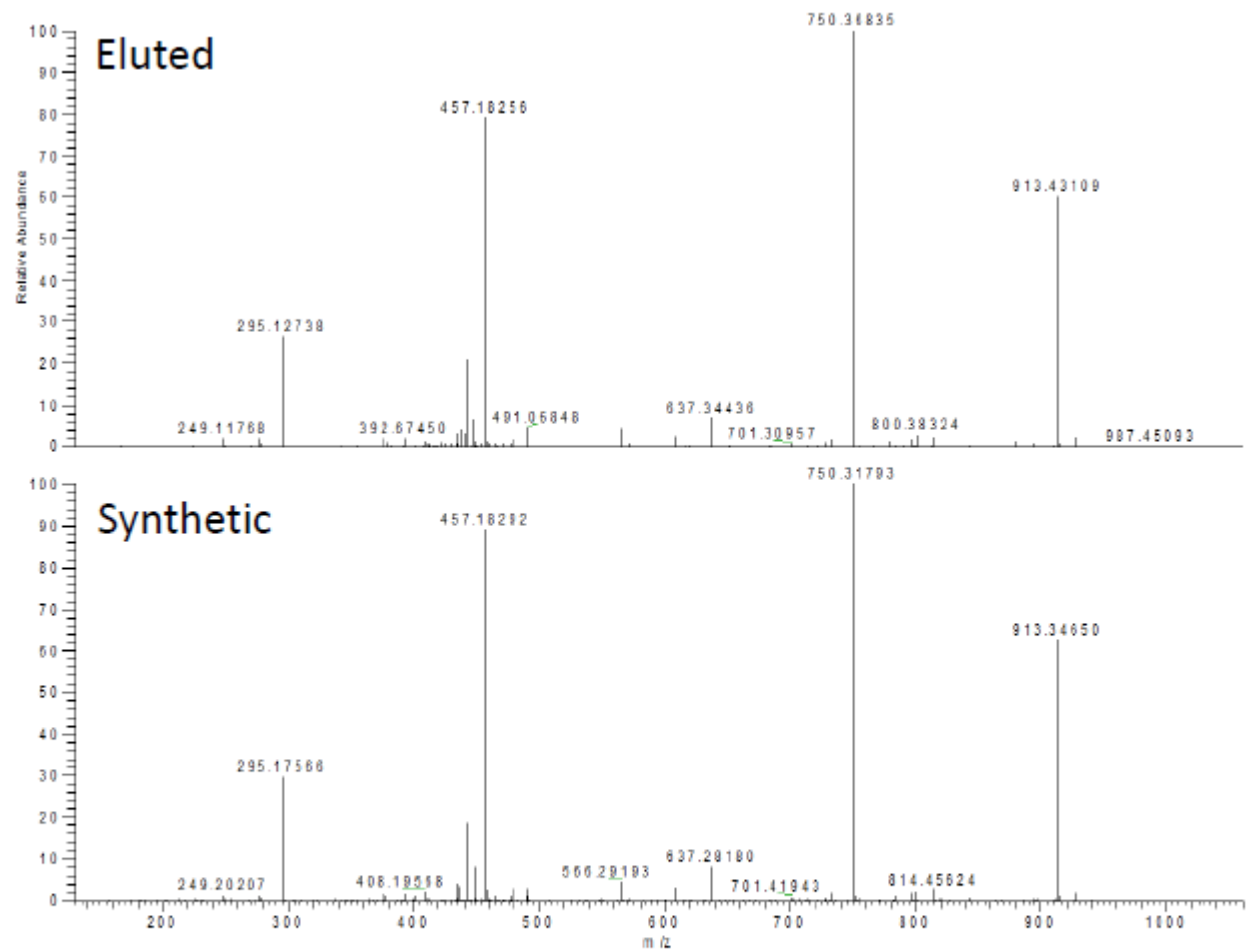
Supplementary Figure 2 B) HLA-B*3801 Epitope PPI₅₋₁₄



b _n ions	132	288.1	401.2	<u>514.3</u>	611.4	<u>724.5</u>	<u>837.5</u>	<u>908.6</u>	<u>1021.7</u>	1134.7
Peptide	M	R	L	L	P	L	L	A	L	L
y _n ions	1177.8	<u>1021.7</u>	865.6	752.5	<u>639.4</u>	542.4	429.3	316.2	<u>245.2</u>	132.1

SUPPLEMENTARY DATA

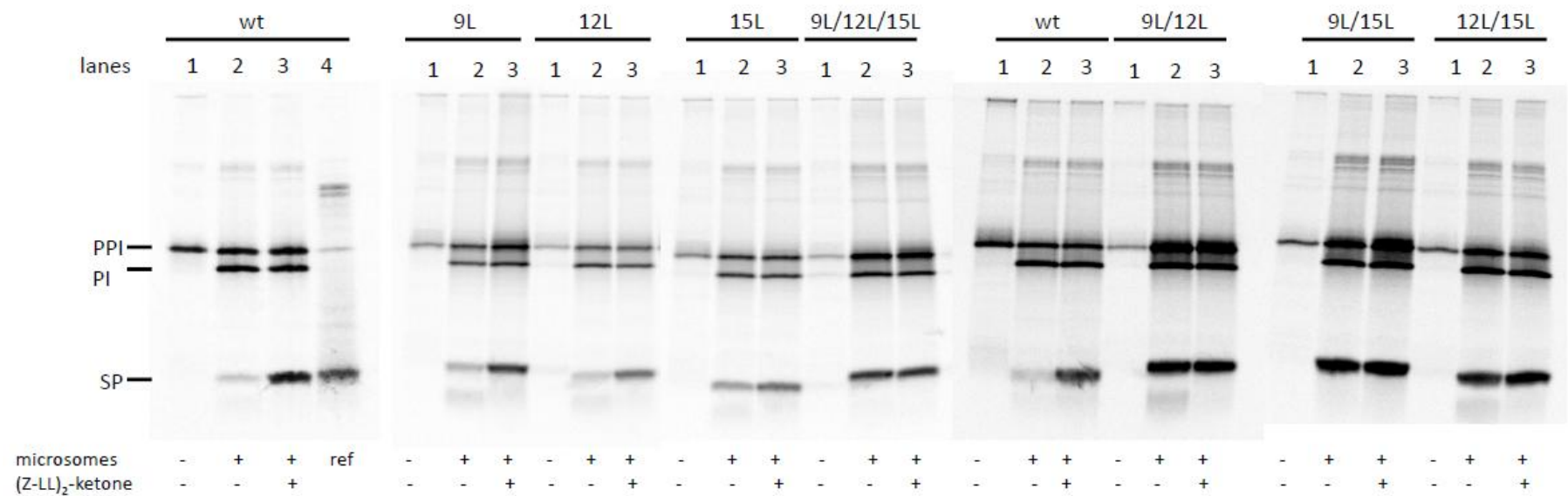
Supplementary Figure 2 C) HLA-B*3801 Epitope PPI₃₃₋₄₁



b _n ions	88	<u>225.1</u>	<u>338.2</u>	<u>437.3</u>	<u>566.3</u>	<u>637.3</u>	<u>750.4</u>	<u>913.5</u>	1044.6
Peptide	S	H	L	V	E	A	L	Y	L
y _n ions	1044.6	957.5	<u>820.5</u>	<u>707.4</u>	<u>608.3</u>	<u>479.3</u>	<u>408.2</u>	<u>295.2</u>	132.1

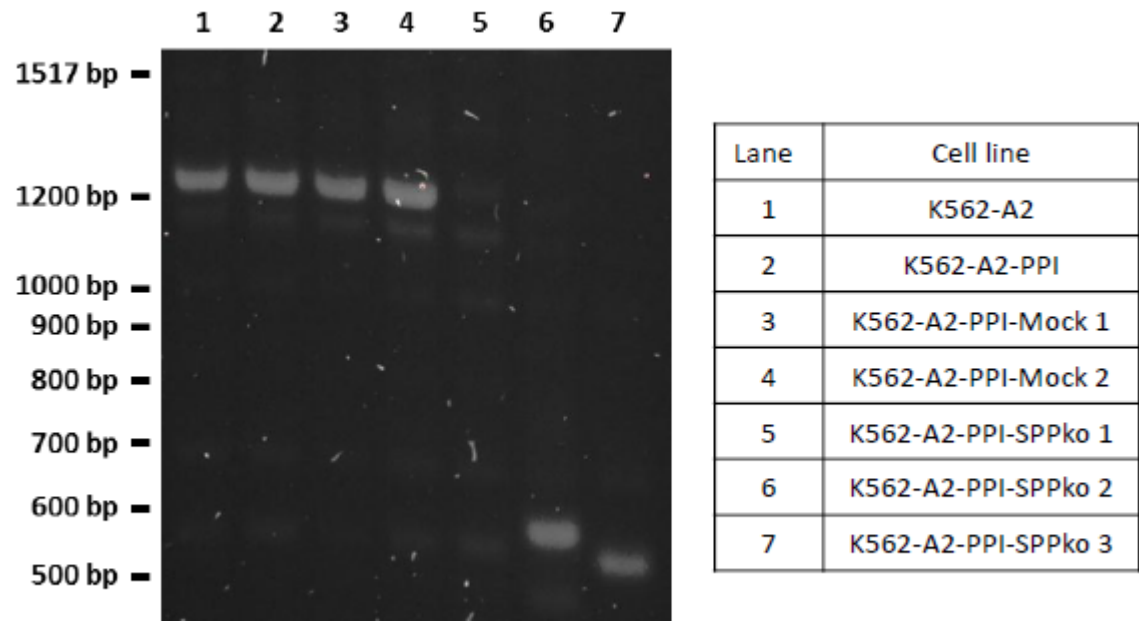
SUPPLEMENTARY DATA

Supplementary Figure S3. Processing of the PPI signal peptide in microsomes. *In vitro* translation of wt PPI mRNA or mutant PPI mRNA (P9L, A12L, A15L) in the absence (lanes 1) or in the presence of ER-derived microsomes (lanes 2 and 3) and SPP inhibitor (Z-LL)₂-ketone (lanes 3). Microsomes were isolated and analyzed by SDS-PAGE, and radiolabeled proteins visualized by phosphorimaging. Lane 4, *in vitro*-translated reference. Images are representative images of n=2, apart from wt where n=5. Equal translocation efficiency and PPI precursor availability for processing was controlled by comparing the amount of proinsulin between conditions.



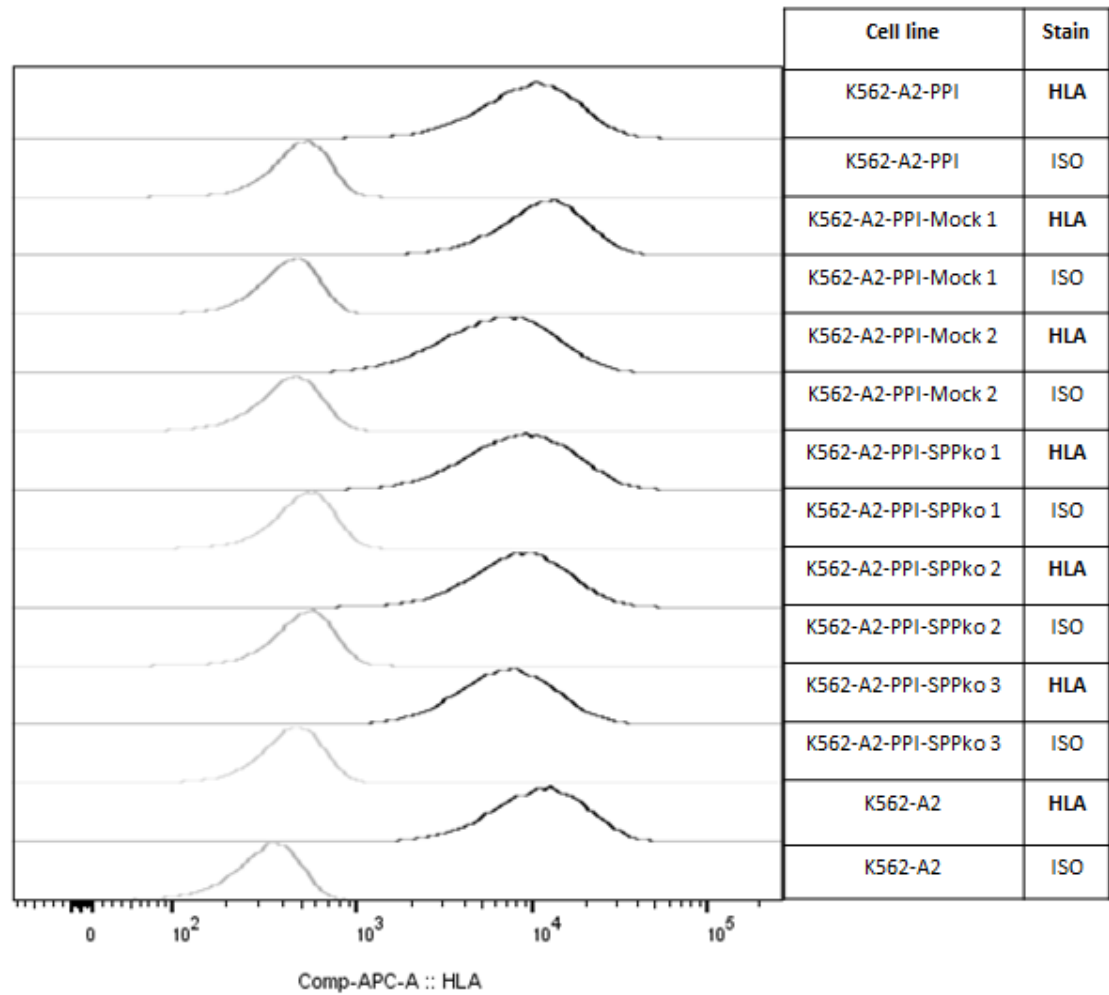
SUPPLEMENTARY DATA

Supplementary Figure S4. Validation of SPP knockout (truncation) by PCR. Within the K562-A2-PPI cells line the SPP gene was truncated using CRISPR-Cas9 directed double targeting of each exon at the start and end of the gene. Simultaneous targeting leads to truncation of the gene in effect leading to functional gene knockout. Length of the gene was assessed using PCR with SPP specific primers on cDNA generated from isolated mRNA from the cell lines. WT (lane 1 and 2) and mock transfected (lane 3 and 4) cell lines harbour full length SPP, whereas reduction (lane 6 and 7) in size or abrogation (lane 5) was observed for cell lines with effective CRISPR-Cas9 targeting of SPP.



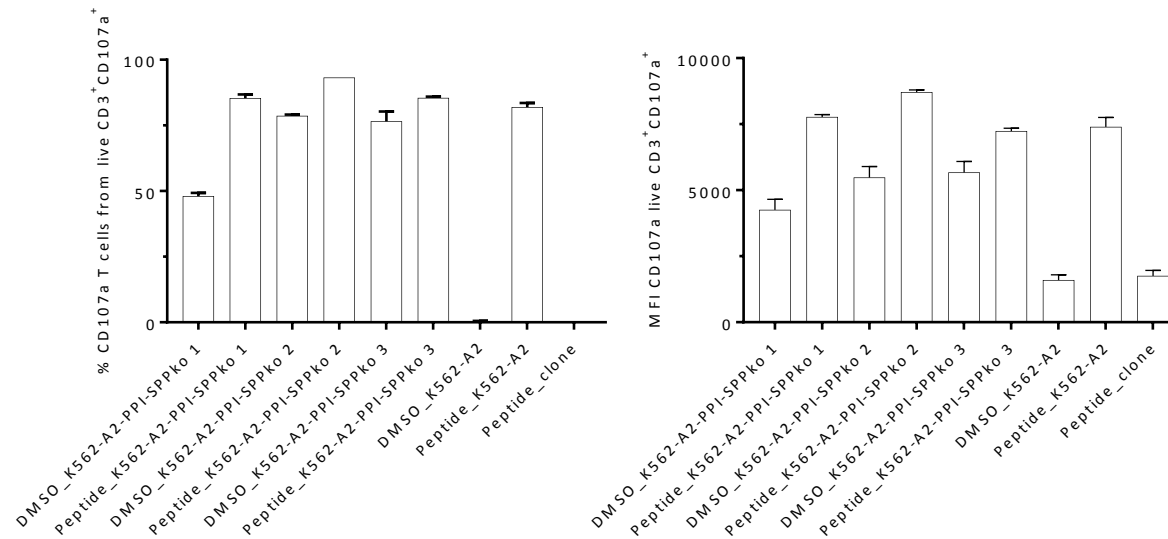
SUPPLEMENTARY DATA

Supplementary Figure S5. HLA class I surface expression on Mock and SPPko cell lines. Cell lines were stained with anti-pan HLA class I antibody (HLA, black line) and corresponding isotype control (ISO, grey line) antibody. Levels of surface HLA expression are similar for each experimental condition (WT, mock and SPP knockout).



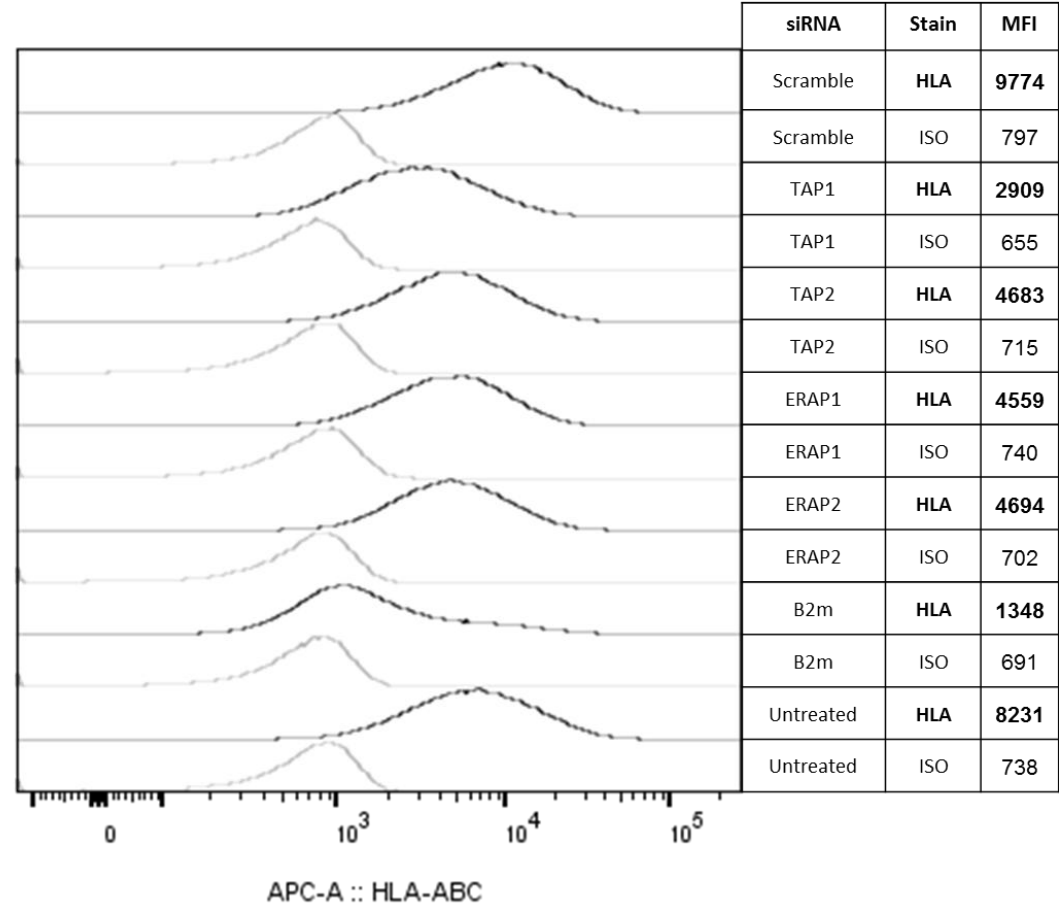
SUPPLEMENTARY DATA

Supplementary Figure S6. Pulsing SPPko cells with cognate peptide rescues their phenotype. K562-A2 and K562-A2-PPI-SPPko cell lines were pulsed for 1 hour with 10uM/ml peptide or peptide diluent (DMSO) prior to co-culture with T cell clone specific for PPI₁₅₋₂₄-HLA-A0201. T cell activation was assessed by CD107a expression. For each SPPko cell line, peptide pulsing leads to increased T cell activation as evidenced by both increases in percentage expression of CD107a (left) and median fluorescence intensity of CD107a (right) with comparable levels in the pulsed WT K562-A2 cell line. Three independent experiments.



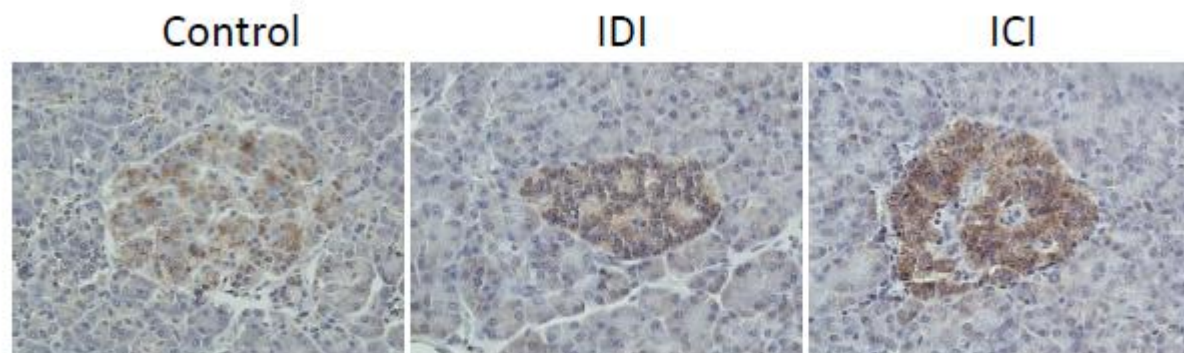
SUPPLEMENTARY DATA

Supplementary Figure S7. HLA class I expression in siRNA experiments. Surface HLA-ABC (W6/32 clone) expression on b2M, ERAP1, ERAP2, TAP1, TAP2, scramble knockdown treated and untreated K562-A24-PPI respectively (HLA, black line) and isotype control (ISO, grey line). Median Fluorescence Intensity (MFI) of HLA staining is shown.



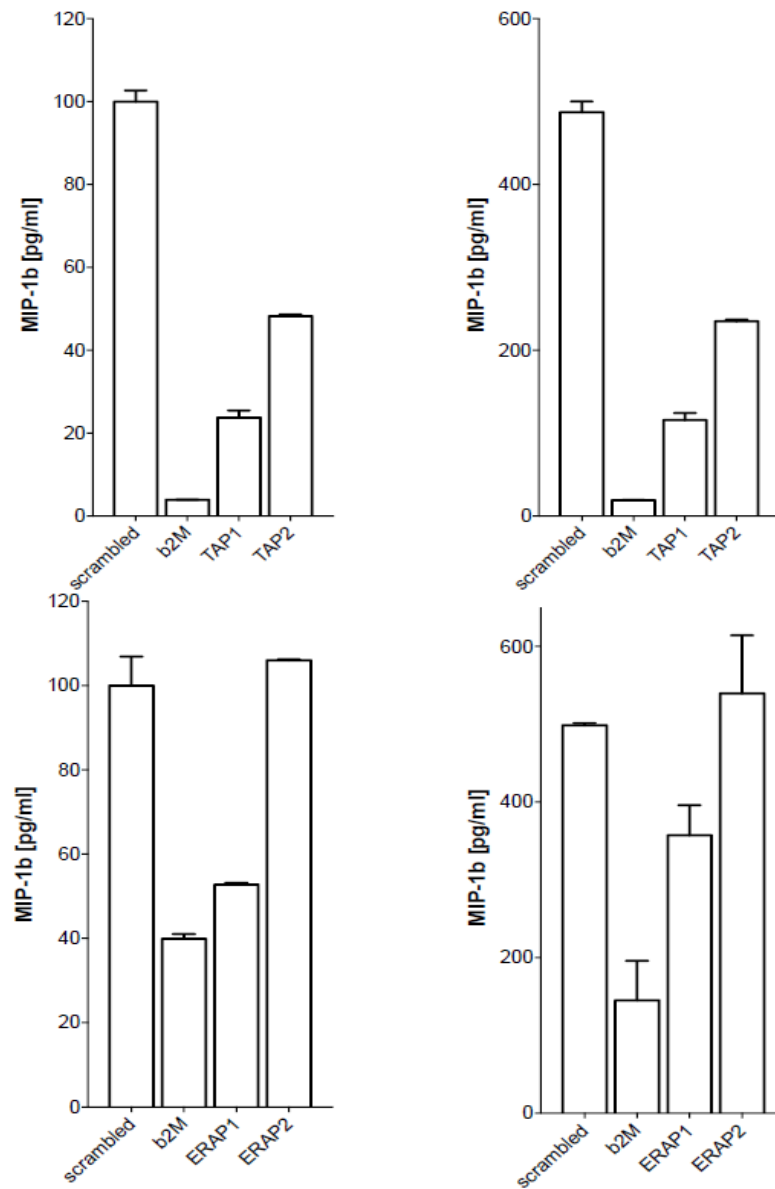
SUPPLEMENTARY DATA

Supplementary Figure S8. Representative immunohistochemistry staining of ERAP. Representative immunohistochemistry staining of ERAP on pancreas samples from patients with type 1 diabetes (n=3) and a representative control sample. ERAP is expressed in all islet cells irrespective of sample source and pancreas samples from type 1 diabetes patients show similar expression in insulin-deficient islets (IDI) and insulin-containing islets (ICI). These data suggest that ERAP expression is not significantly altered in the islets of patients with type 1 diabetes.



Supplementary Figure S9. MIP-1 β in siRNA experiments. MIP-1 β production of HLA-A2402-restricted PPI₃₋₁₁ specific CD8 T cell clone 4C6 upon co-culture with *β 2M*, *TAP1*, *TAP2*, genes (top panel) and *ERAP1*, *ERAP2* gene (bottom panel) knockdown in K562-A24-PPI cells. Two independent experiments.

SUPPLEMENTARY DATA



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References

1. Eichmann M, de Ru A, van Veelen PA, Peakman M, Kronenberg-Versteeg D: Identification and characterisation of peptide binding motifs of six autoimmune disease-associated human leukocyte antigen-class I molecules including HLA-B*39:06. *Tissue antigens* 2014;84:378-388
2. The UniProt C: UniProt: the universal protein knowledgebase. *Nucleic Acids Res* 2017;45:D158-D169
3. Rammensee H, Bachmann J, Emmerich NP, Bajor OA, Stevanovic S: SYFPEITHI: database for MHC ligands and peptide motifs. *Immunogenetics* 1999;50:213-219