

SUPPLEMENTARY DATA

**Supplementary Table 1.** Primary antibodies for Western blot.

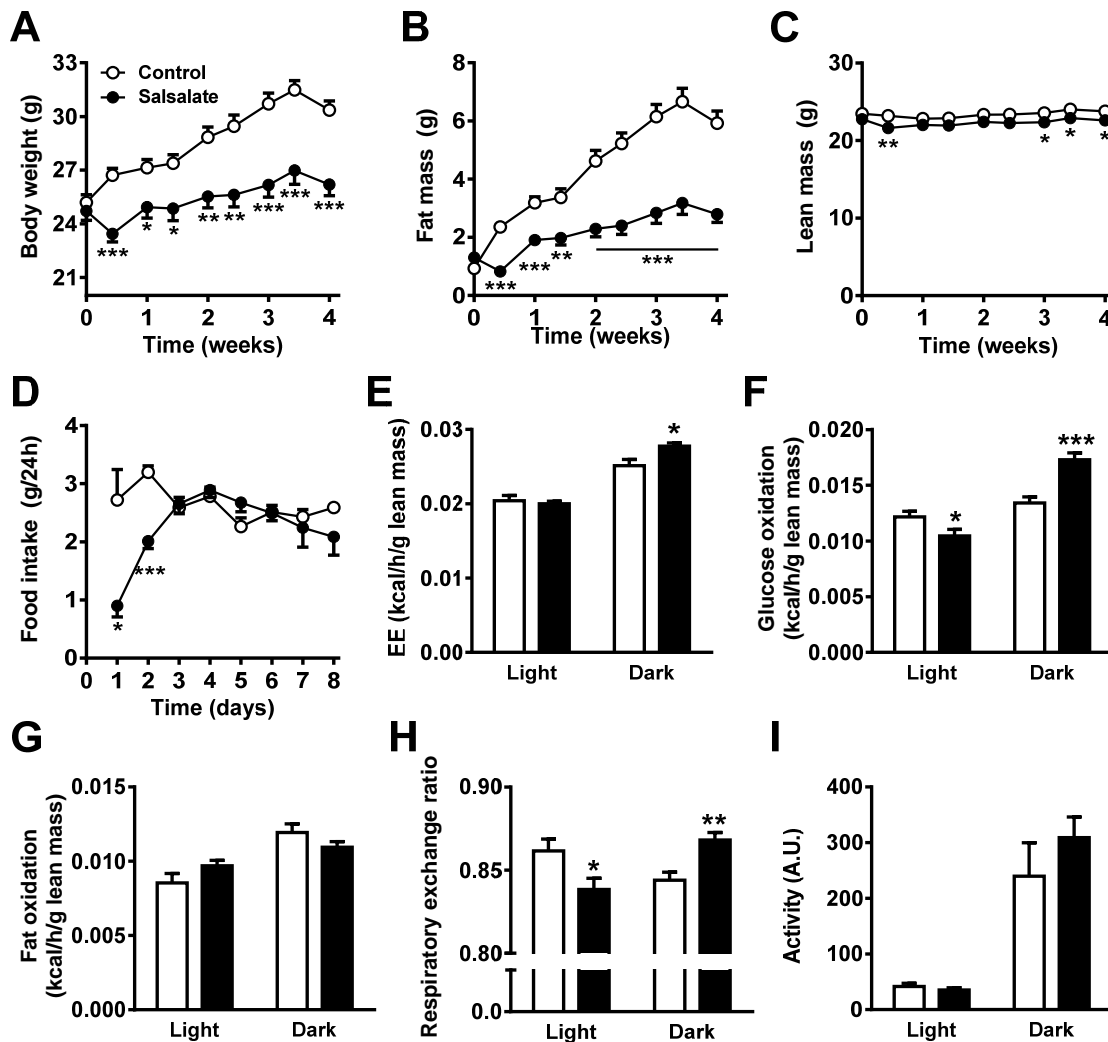
Primary antibody	Residue	Supplier	Reference	Dilution
ACC	-	Cell Signaling	#3662	1:1000
pACC	Ser79	Cell Signaling	#3661	1:1000
AMPK $\alpha$	-	Cell Signaling	#2532	1:1000
pAMPK $\alpha$	Thr172	Cell Signaling	#2535	1:1000
pHSL	Ser563	Cell Signaling	#4139	1:1000
pHSL	Ser565	Cell Signaling	#4137	1:1000
PKA substrates	Ser/Thr	Cell Signaling	#9621	1:1000
$\alpha/\beta$ Tubulin	-	Cell Signaling	#2148	1:1000

**Supplementary Table 2.** Primer sequences of forward and reverse primers (5'  $\rightarrow$  3').

Gene	Forward primer	Reverse Primer
<i>36b4</i>	GGACCCGAGAAGACCTCCTT	GCACATCACTCAGAATTTCAATGG
<i>Acc1</i>	AACGTGCAATCCGATTTGTT	GAGCAGTTCTGGGAGTTTCG
<i>Acc2</i>	AGATGGCCGATCAGTACGTC	GGGGACCTAGGAAAGCAATC
<i>Adrb3</i>	TGAAACAGCAGACAGGGACA	AGTCTGTCAGCTTCCCTCCA
<i>Cd36</i>	GCAAAGAACAGCAGCAAATC	CAGTGAAGGCTCAAAGATGG
<i>Cpt1a</i>	AGGAGACAAGAACCCCAACA	AAGGAATGCAGGTCCACATC
<i>Elovl3</i>	TGTTGGCCAGACCTACATGA	ATCCGTGTAGATGGCAAAGC
<i>F4/80</i>	CTTTGGCTATGGGCTTCCAGTC	GCAAGGAGGACAGAGTTTATCGTG
<i>Fasn</i>	CACAGGCATCAATGTCAACC	TTTGGGAAGTCCTCAGCAAC
<i>Fizz1</i>	CCTGCCCTGCTGGGATGACT	GGGCAGTGGTCCAGTCAACGA
<i>Mcp1</i>	GCATCTGCCCTAAGGTCTTCA	TTCACTGTCACTGGTCACTCCTA
<i>Nos2</i>	TCCTGGACATTACGACCCCT	CTCTGAGGGCTGACACAAGG
<i>Ppargc1a</i>	TGCTAGCGGTTCTCACAGAG	AGTGCTAAGACCGCTGCATT
<i>Ppara</i>	CAACCCGCCTTTTGTTCATAC	CCTCTGCCTCTTTGTCTTCG
<i>Scd1</i>	GCTCTACACCTGCCTCTTCGGGAT	TCCAGAGGCGATGAGCCCCG
<i>Srebp1c</i>	CTGGCTGAGGCGGGATGA	TACGGGCCACAAGAAGTAGA
<i>Ucp1</i>	TCAGGATTGGCCTCTACGAC	TGCATTCTGACCTTCACGAC
<i>Ym1</i>	ACAATTAGTACTGGCCCACCAGGAA	TCCTTGAGCCACTGAGCCTTCA
$\beta$ 2m	TGACCGGCTTGTATGCTATC	CAGTGTGAGCCAGGATATA

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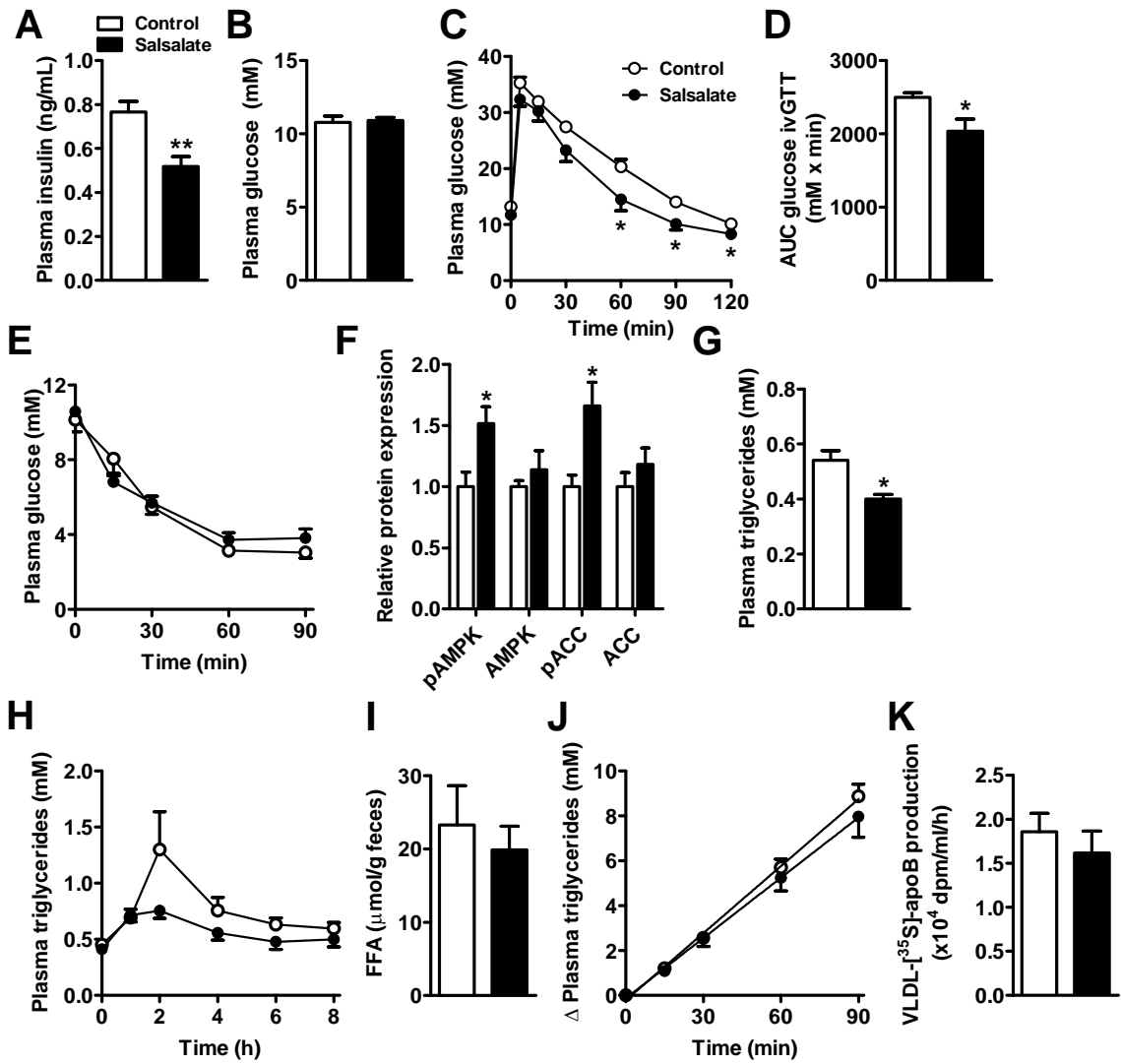
**Supplementary Figure 1. Salsalate prevents obesity and fat mass accumulation in wild type mice fed a high fat diet.** 10-week old male wild type mice (C57Bl/6J background) were fed a high fat diet (HFD) without (open circles) or with (closed circles) salsalate for 4 weeks (A-D). Body weight (A), fat mass (B) and lean mass (C) were monitored throughout the experiment. Food intake was measured daily in the first week of the experiment (D). From day 5 to day 9 of salsalate treatment, mice were housed in fully automatic metabolic cages (LabMaster System; TSE Systems, Bad Homburg, Germany), which measured oxygen uptake ( $V_{O_2}$ ), carbon dioxide production ( $V_{CO_2}$ ) and caloric intake. Total energy expenditure (E) was calculated from  $V_{O_2}$  and  $V_{CO_2}$  using the Weir equation, and glucose oxidation (F) and fat oxidation (G) were calculated from  $V_{O_2}$  and  $V_{CO_2}$  as described previously (1). Respiratory exchange ratio (H) was also calculated from  $V_{O_2}$  and  $V_{CO_2}$ . Physical activity (I) was measured with infrared sensor frames. Measurements (E-H) were corrected for lean mass as determined by EchoMRI (EchoMRI-100, Houston, Texas, USA). EE = energy expenditure. Values represent means  $\pm$  SEM (n=7-8). \*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$  vs control.



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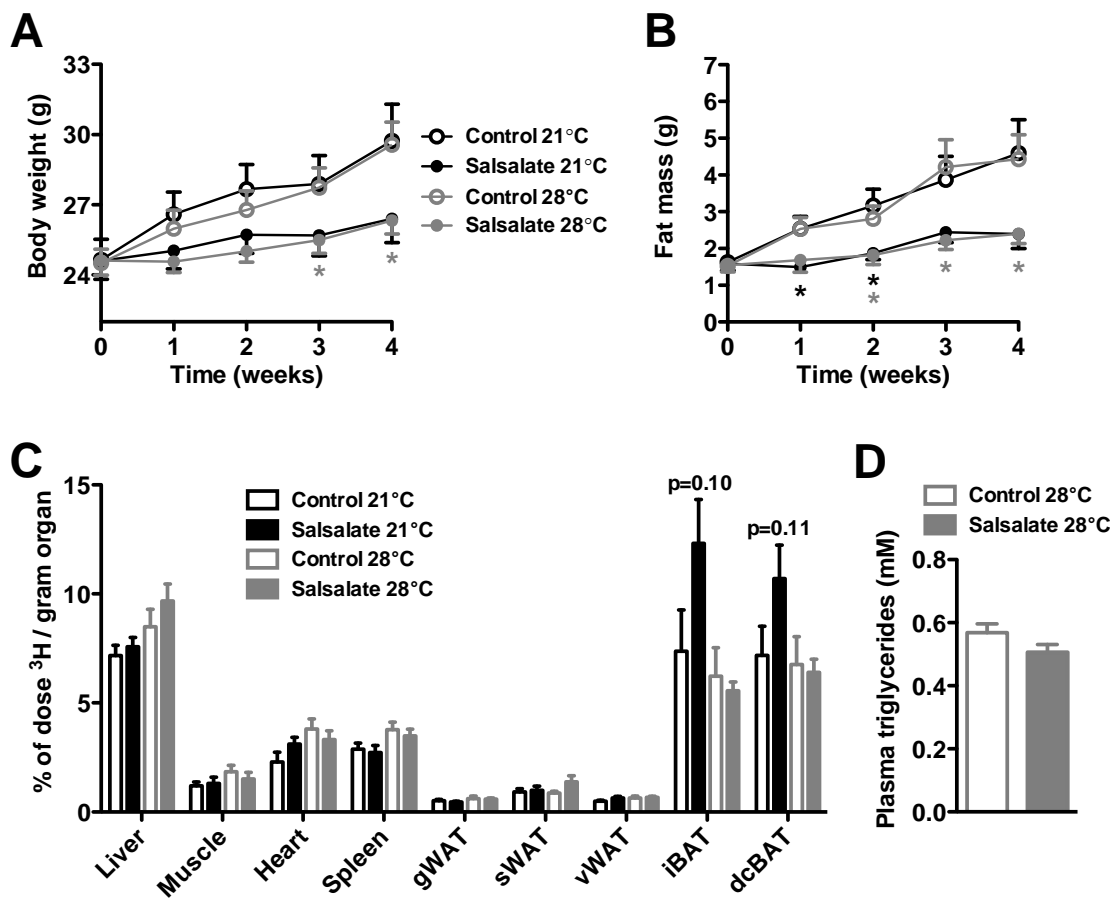
**Supplementary Figure 2. Salsalate improves glucose and triglyceride metabolism in wild type mice fed a high fat diet.** 10-week old male wild type mice were fed a high fat diet (HFD) without (open circles/bars) or with (closed circles/bars) salsalate for 4 weeks. Blood samples from 6 h-fasted mice were collected by tail bleeding at different time points and plasma insulin (A), glucose (B) and triglyceride (G) levels were determined. After 4 weeks, 6 h fasted mice were i.v. injected with glucose and additional blood samples were taken at 5, 15, 30, 60, 90 and 120 min after injection (C-D). After 4 weeks, an insulin tolerance test was performed by injecting 6 h fasted mice i.p. with insulin (NovoRapid, Novo Nordisk, Denmark; 1 U/kg whole body mass) and measuring glucose at  $t = 15, 30, 60$  and  $90$  min (E). Protein content in muscle was determined by Western blot (F). Postprandial triglyceride (TG) response was measured after 3 weeks of salsalate treatment. Animals were fasted for 6 h and a basal blood sample was drawn before an intragastric load of 200  $\mu$ L olive oil (Carbonell, Traditional, Cordoba, Spain) was given. Blood samples were drawn 1, 2, 4, 6 and 8 h after the bolus via tail vein bleeding and plasma TG levels were measured (H). Feces was collected during the 3rd week of salsalate treatment. Feces was then weighed, freeze-dried and ground, and fecal fatty acids were determined by methyl esterification. To this end, 750  $\mu$ L  $\text{CH}_3\text{OH}/\text{NaOH}$  (10 M)/ $\text{H}_2\text{O}$  (3:1:1 v/v) was added to the feces and samples were incubated and mixed in a thermomixer (600 rpm,  $90^\circ\text{C}$ ) for 1 h. Then, 1050  $\mu$ L of HCl (6 M)/hexane (3:7.7 v/v) was added before samples were vortexed and spun down 1 min at 14,000 rpm. The upper hexane layer was dried using  $\text{N}_2$  and redissolved in 2% Triton X-100. Fatty acids were measured using the NEFA C kit (Wako Diagnostics, Instruchemie, Delfzijl) (I). After 4 weeks of salsalate treatment, hepatic VLDL production was determined. Mice were fasted for 4 h and anesthetized by intraperitoneal injection of 6.25 mg/kg acepromazine (Alfasan, Weesp), 6.25 mg/kg midazolam (Roche, Mijdrecht), and 0.31 mg/kg fentanyl (Janssen Pharmaceuticals, Tilburg). Mice were injected intravenously with Tran<sup>[35S]</sup> label (20  $\mu$ Ci/mouse; MP Biomedicals, Eindhoven) to label newly produced apolipoprotein B (apoB). After 30 min, at  $t = 0$  min, Triton WR-1339 (Sigma-Aldrich) was injected intravenously (0.5 mg/g body weight, 10% solution in PBS) to block serum VLDL clearance. Blood samples were drawn before ( $t = 0$ ) and at 15, 30, 60, and 90 min after injection of Triton and used for determination of plasma TG concentration (J). After 120 min, mice were exsanguinated via the retro-orbital plexus. VLDL was isolated from serum after density gradient ultracentrifugation at  $d < 1.006$  g/ml by aspiration (2) and examined for incorporated <sup>35</sup>S-activity as a measure of ApoB production rate (K). AUC = area under the curve; FFA = free fatty acids. Values represent means  $\pm$  SEM (n=5-8). \*  $p < 0.05$ , \*\*  $p < 0.01$  vs control.

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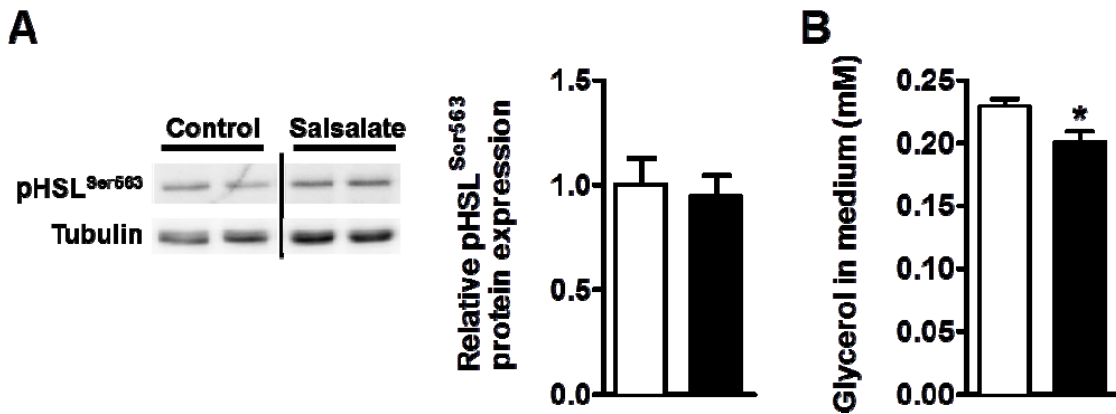
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**Supplementary Figure 3. Effect of salsalate at thermoneutrality** 10-week old male wild type mice were housed at 21°C (black circles/bars) or 28°C (grey circles/bars) and fed a high fat diet (HFD) without (open circles/bars) or with (closed circles/bars) salsalate for 4 weeks. Body weight (A) and fat mass (B) were monitored throughout the experiment. A clearance experiment was performed in which 6 h-fasted mice were i.v. injected with [<sup>3</sup>H]TO-labeled lipoprotein-like emulsion particles. After 15 min, mice were sacrificed and uptake of [<sup>3</sup>H]TO-derived activity was determined in the organs (C). After 4 weeks of treatment, blood samples from 6 h-fasted mice were collected by tail bleeding and plasma triglyceride levels were determined (D). Values represent means ± SEM (n=7-8). \* p<0.05 vs control. (*g,s,v*)WAT, gonadal, subcutaneous, visceral white adipose tissue; (*i,dc*)BAT, interscapular, dorsocervical brown adipose tissue.

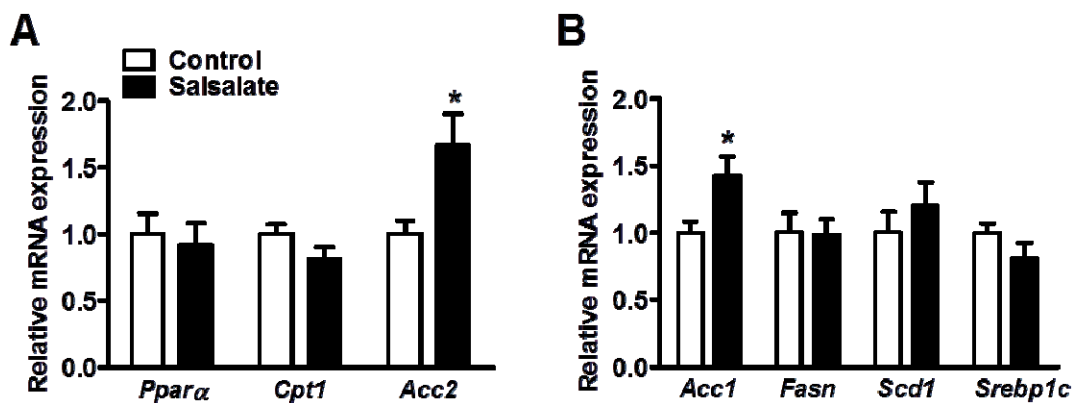


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**Supplementary Figure 4. Effect of salsalate on 3T3-L1 cells.** 3T3-L1 cells were cultured and differentiated into white adipocytes. White adipocytes were treated with 300  $\mu$ M salsalate for 8 h. Protein content was determined by Western blot (A) and glycerol concentration in the culture medium was measured (B). Values represent means  $\pm$  SD (n=3). \*  $p < 0.05$  vs control.



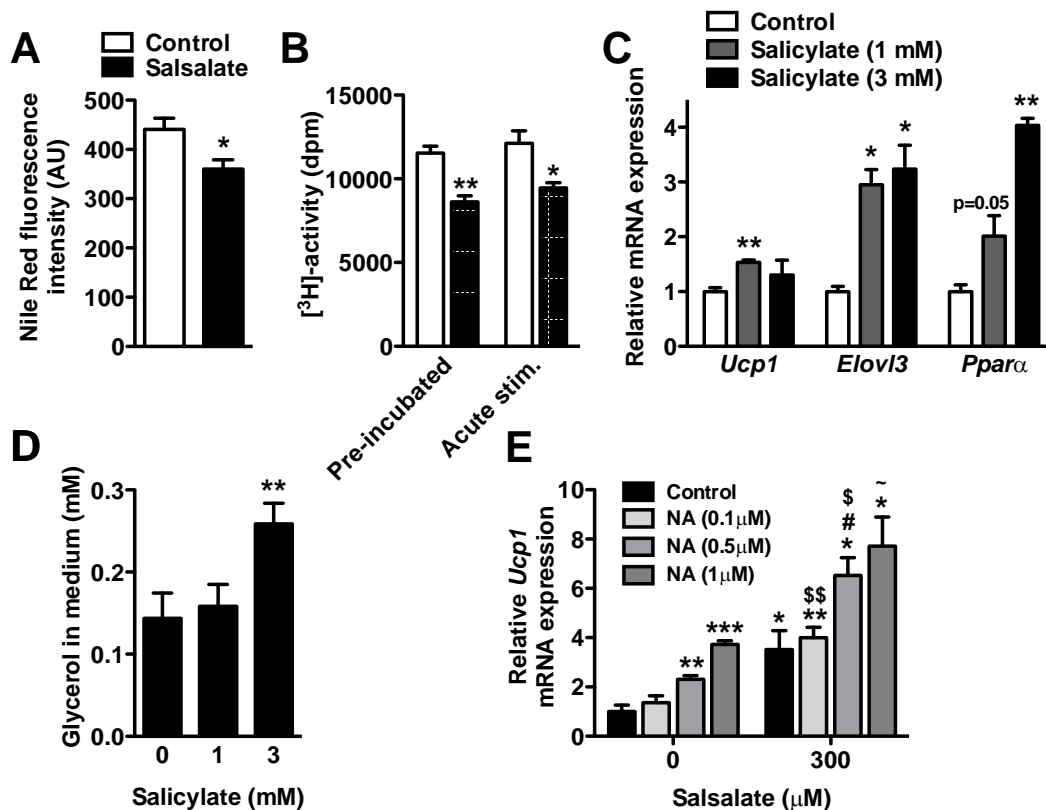
**Supplementary Figure 5. Effect of salsalate on genes involved in beta-oxidation and lipogenesis in gWAT of wild type mice fed a high fat diet.** 10-week old male wild type mice were fed a high fat diet (HFD) without (open bars) or with (closed bars) salsalate for 4 weeks and gonadal white adipose tissue (gWAT) was collected. mRNA expression was determined by qRT-PCR (A-B). Values represent means  $\pm$  SEM (n=7-8). \*  $p < 0.05$  vs control.



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**Supplementary Figure 6. *In vitro* effects of salsalate and salicylate on T37i brown adipocytes.**

T37i cells were cultured and differentiated into mature brown adipocytes. Cells (grown in 0.32 cm<sup>2</sup> wells) were treated with salsalate (300 μM) for 8 h, stained with 0.1 mg/mL Nile Red solution (Molecular probes, N-1142) for 2 h at 37°C and measured with a fluorimeter (Ex. 485 nm, Em. 590 nm). Values represent means ± SEM of 11 independent wells (A). To assess *de novo* lipogenesis, cells (grown in 3.8 cm<sup>2</sup> wells) were incubated at 37°C with HEPES buffer (29.8 g HEPES, 8.78 g NaCl, 4.66 g KCl, 1.12 g D-glucose, 18.8 g BSA, 0.132 g CaCl<sub>2</sub> in 1.125 L H<sub>2</sub>O, pH 7.3), 100 nM insulin and [<sup>3</sup>H]glucose (~500,000 cpm). Cells were either pre-incubated with vehicle or salsalate for 6 h or the compound was added together with the [<sup>3</sup>H]glucose. After 2 h, cells were washed and 600 μL cold CHCl<sub>3</sub>/CH<sub>3</sub>OH (1:1 v/v) was added. Cells were left on ice for 30 min, upon which the supernatant was isolated. Next, 125 μL H<sub>2</sub>O was added and the samples were mixed and spun for 10 min at 3,500 rpm. The CHCl<sub>3</sub> layer was isolated and evaporated under N<sub>2</sub> and incorporation of [<sup>3</sup>H]glucose-derived radioactivity was quantified. Values represent means ± SEM of 4-5 independent wells (B). Cells were treated with salicylate (1 or 3 mM) for 8 h and mRNA expression was determined by qRT-PCR (C). In addition, glycerol concentration in the medium was assessed (D). Cells were treated with salsalate (300 μM) in the presence of increasing concentrations of noradrenalin (0.1 – 1 μM) for 8 h and *Ucp1* mRNA expression was determined (E). NA = noradrenalin. Values represent means ± SD of 3-4 independent sets of RNA. \* p<0.05, \*\* p<0.01 vs control, \$ p<0.05, \$\$ p<0.01 vs NA, ~ p<0.10 vs NA and salsalate.



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### References

1. van Klinken,JB, van den Berg,SA, Havekes,LM, Willems van,DK: Estimation of activity related energy expenditure and resting metabolic rate in freely moving mice from indirect calorimetry data. *PLoS One* 7:e36162, 2012
2. Redgrave,TG, Roberts,DC, West,CE: Separation of plasma lipoproteins by density-gradient ultracentrifugation. *Anal Biochem* 65:42-49, 1975