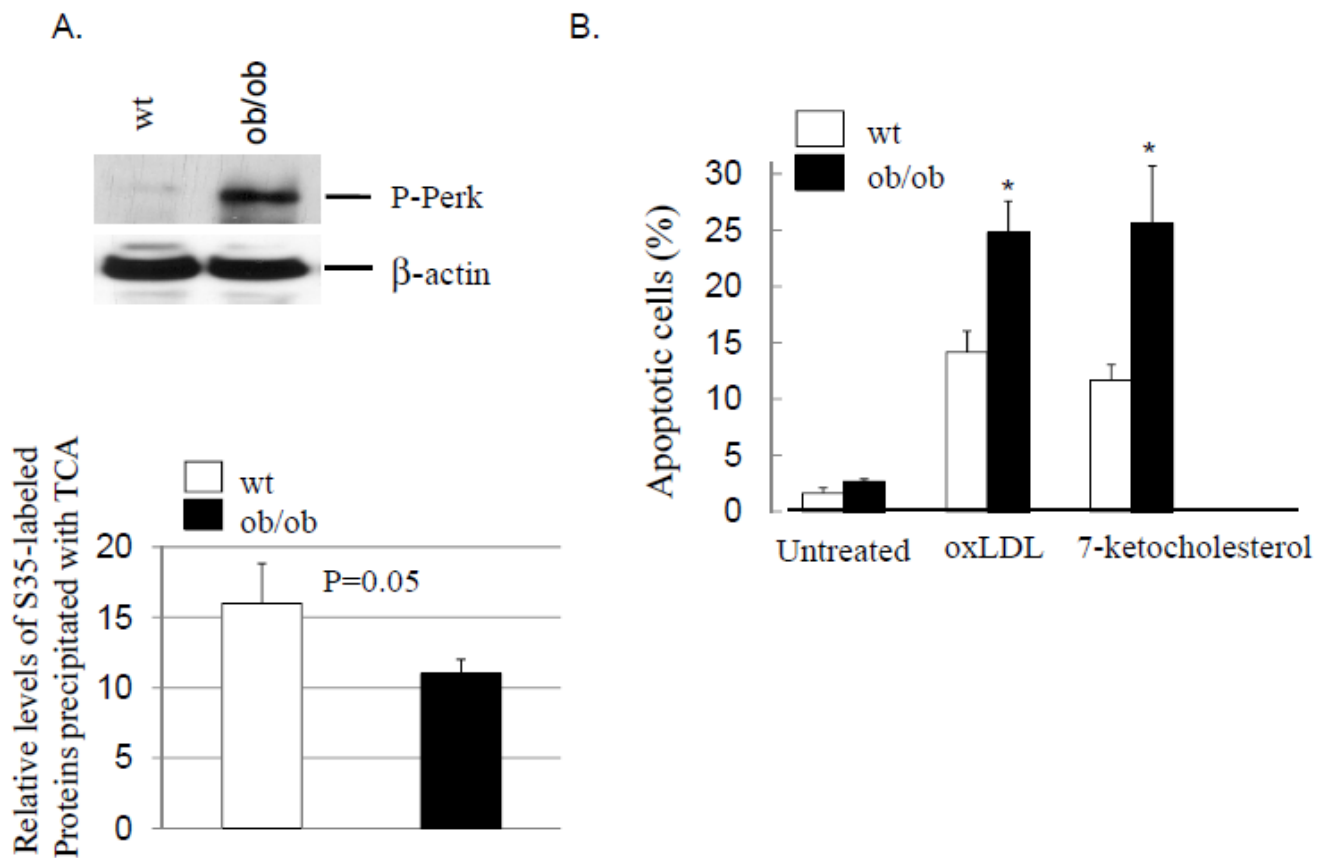


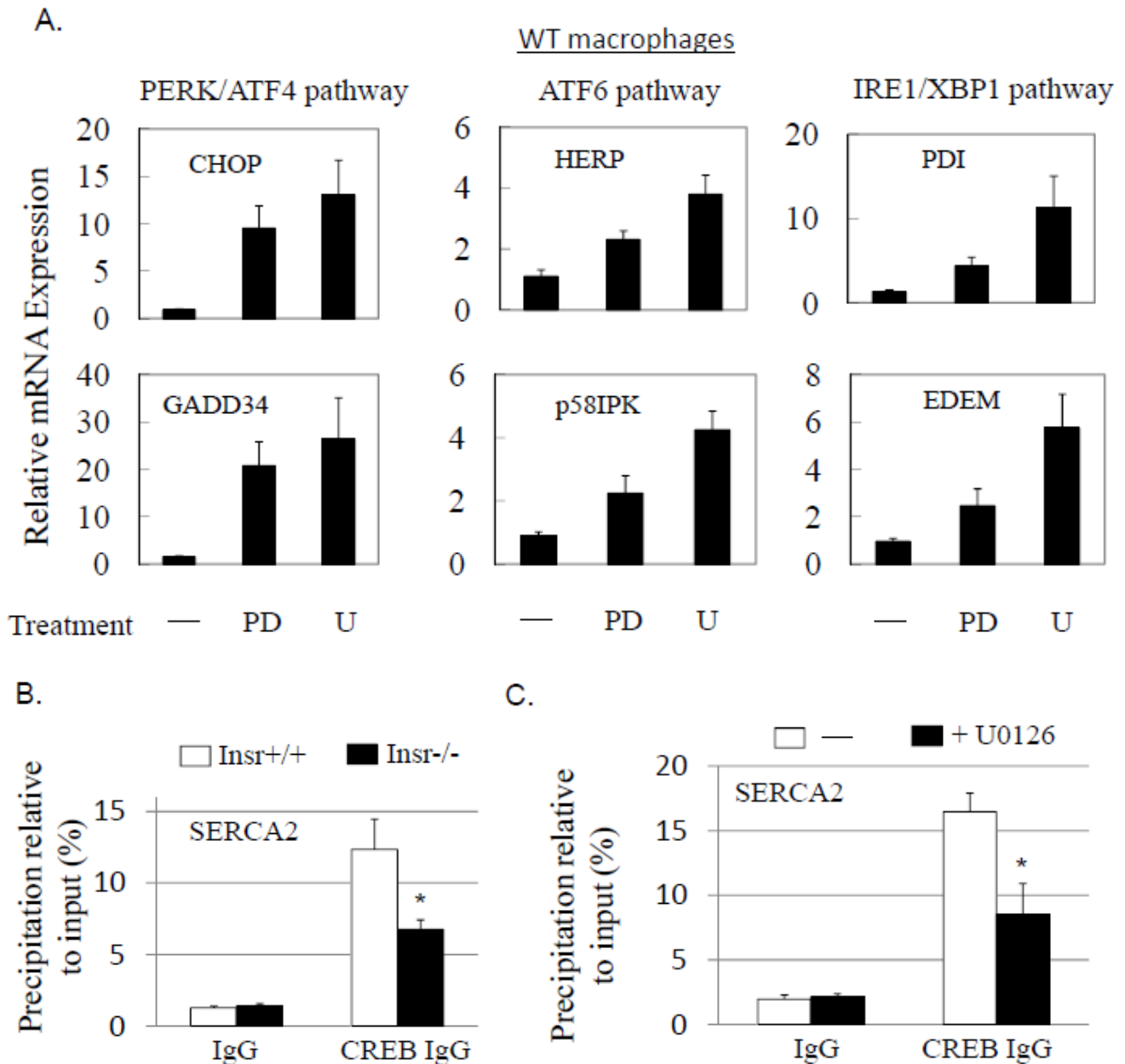
SUPPLEMENTARY DATA

**Supplementary Figure 1.** (A) (Top) Fresh peritoneal macrophages from either ob/ob or lean wild-type (wt) mice fed regular chow diet were cultured at 37°C in DMEM with 10% FBS for 2 hr. Cells were harvested and protein extracts were prepared. Western analysis was performed with antibodies against the proteins as indicated. n=3. (Bottom) Pulse-labeling of cells from wt and ob/ob mice with 35S-methionine was performed. The amounts of 35S label incorporation by trichloroacetic acid (TCA) precipitation were then measured. The results were normalized to total cell counts. (B) Pooled ConA-elicited peritoneal macrophages were incubated with or without oxLDL (100 µg/ml) or 7-ketocholesterol (40 µg/ml) at 37°C for 8 h. Apoptosis of macrophages was determined by annexin V staining. All results represent average ± SE. \*, P<0.05 for oxLDL or 7-ketocholesterol-loaded wt vs. ob/ob cells. n=3. (C) ConA-elicited peritoneal macrophages isolated from ob/ob and wt mice were treated as described in Fig 1A. The levels of indicated mRNAs regulated by PERK/ATF4, ATF6, and IRE1/XBP1 arms of the UPR were measured by real time QPCR. n=3.



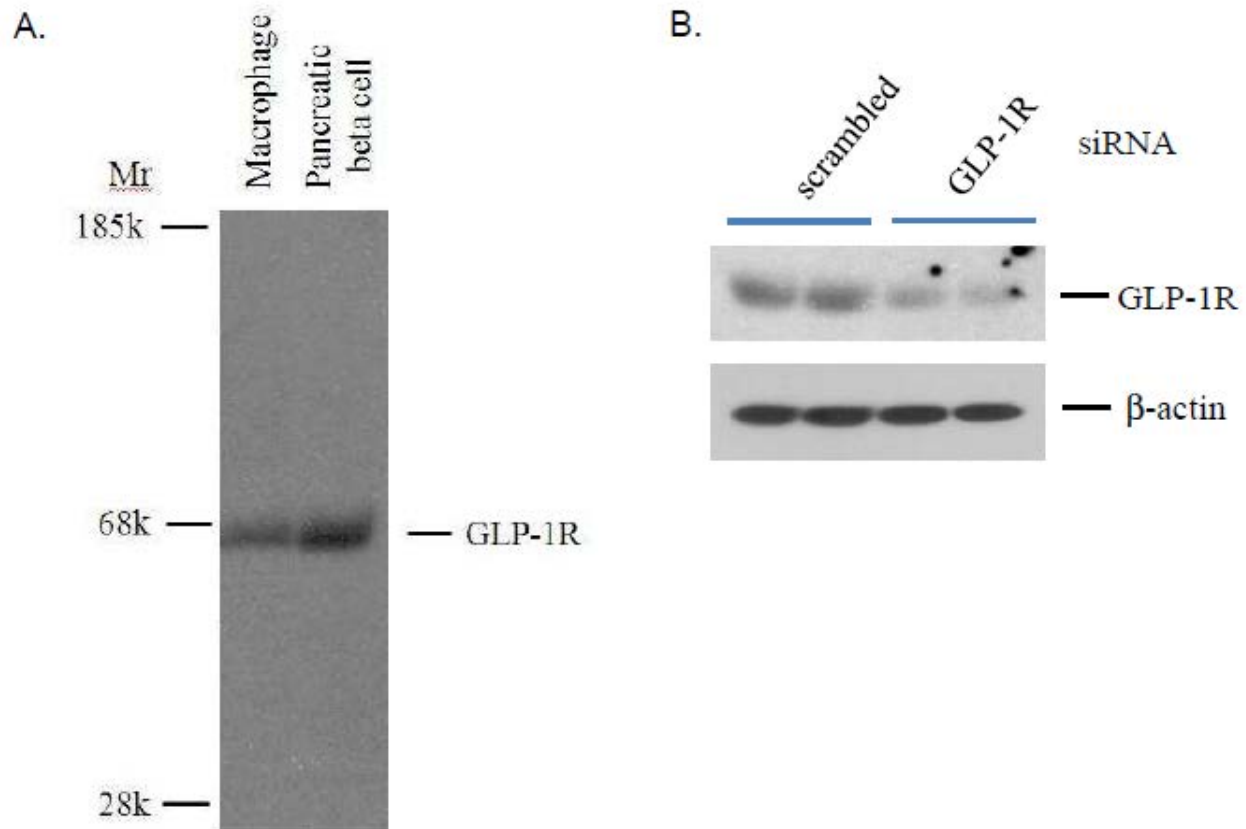
SUPPLEMENTARY DATA

**Supplementary Figure 2.** (A) The mRNA expression of macrophage PERK/ATF4, ATF6, and IRE1/XBP1 target genes was measured by real time QPCR as described in Fig 1A. QPCR was performed in triplicate. n=3. (B) Chromatin immunoprecipitation (ChIP) analysis of the association of P-CREB to SERCA2 gene promoter in *Insr*<sup>+/+</sup> and *Insr*<sup>-/-</sup> macrophages. Cells were cultured as described in (A), and ChIP analysis was performed with a ChIP assay kit. ChIP samples were quantified by real-time qPCR and results were presented as percent precipitation relative to input chromatin. n=4. \*,P<0.05. (C) ChIP analysis of the association of P-CREB to SERCA2 gene promoter in wild-type macrophages with or without 4h-treatment of MEK inhibitor U0126 (10 uM). ChIP analysis was performed with a ChIP assay kit. ChIP samples were quantified by real-time qPCR and results were presented as percent precipitation relative to input chromatin. n=3. \*,P<0.05.

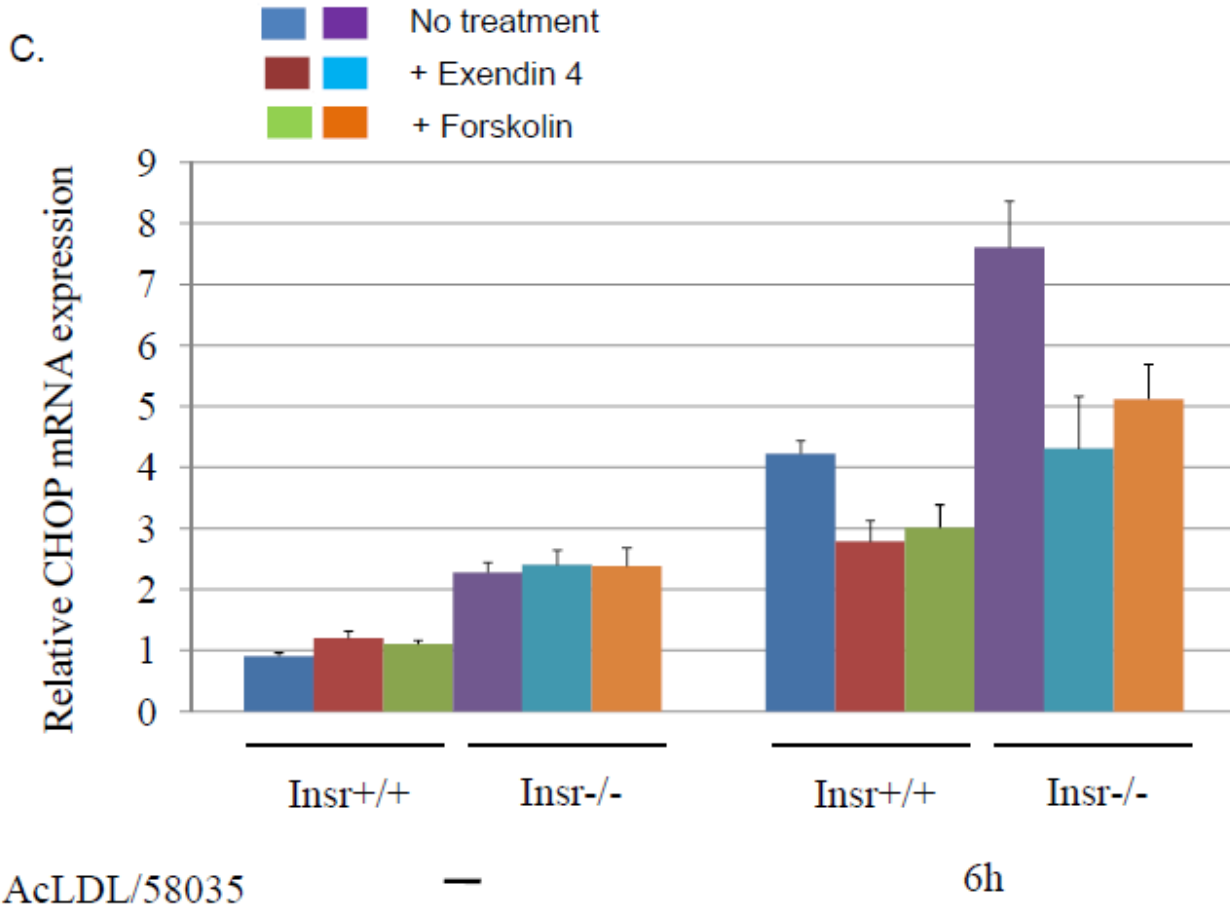


SUPPLEMENTARY DATA

**Supplementary Figure 3.** (A) The expression of GLP-1 receptor in primary macrophages from wild-type mice as well as in MIN6 mouse pancreatic beta cells was determined by Western analysis using anti-GLP-1 receptor antibody. (B) Mouse primary macrophages were transfected with 100 nM scrambled (non-targeting) or GLP-1 receptor (GLP-1R) siRNAs for 2 days. Protein expression of GLP-1 receptor and actin was measured by Western analysis. (C) *Insr*<sup>+/+</sup> or *Insr*<sup>-/-</sup> macrophages were treated with AcLDL and compound 58035 with or without exendin-4 (100 nM) or forskolin (10  $\mu$ M) for indicated times. CHOP mRNA expression was monitored by real time QPCR. QPCR was performed in triplicate. n=3.

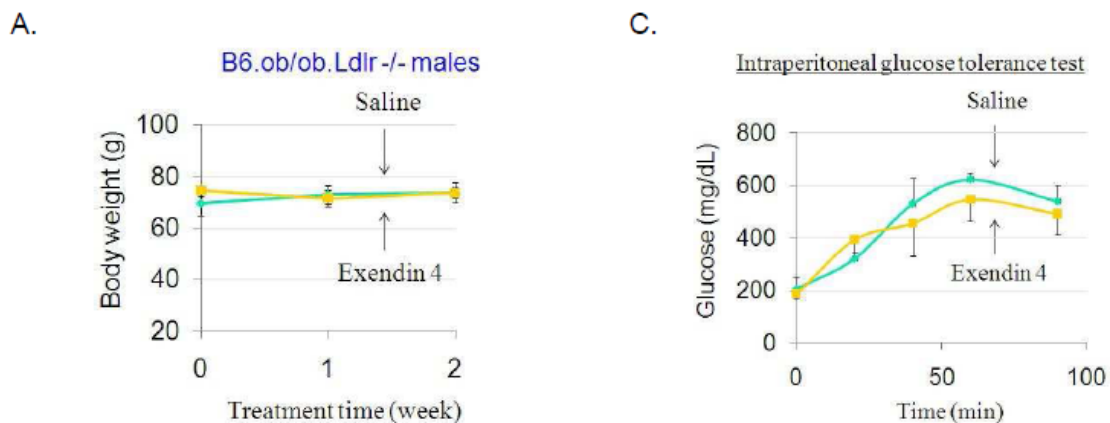


SUPPLEMENTARY DATA



SUPPLEMENTARY DATA

**Supplementary Figure 4.** Characterization of *Ldlr*<sup>-/-</sup> and *ob/ob*;*Ldlr*<sup>-/-</sup> mice treated with exenatide or saline. **(A)** Body weight of mice during the period of two-week treatment. Mice were fed Western-type diet for 3 months then received either PBS or exendin 4 (20 ng/g body weight) for two weeks under pair-feeding condition. n=4 for each *ob/ob*;*Ldlr*<sup>-/-</sup> group. **(B)** Plasma metabolic characteristics of *ob/ob*;*Ldlr*<sup>-/-</sup> mice with PBS or exendin 4 treatment described in (A). Metabolic profiles were monitored with plasma from mice fasted for overnight. **(C)** Glucose tolerance test with mice described in (A). Blood samples were obtained at indicated times after intraperitoneal injection of 2 g/kg body weight dextrose after overnight fasting. Blood glucose was determined using a Accu-Chek glucose monitor. **(D)** Primary macrophages isolated from Western-type diet-fed *ob/ob*;*Ldlr*<sup>-/-</sup> mice receiving PBS or exendin-4 in vivo were used for the analysis of SERCA2 by Western analysis (Left) or of *Xbp1* mRNA splicing by real time QPCR (Right). \*, P<0.05 for mice with in vivo treatment of PBS vs. exendin-4. n=3. **(E)** (Left) ConA-elicited peritoneal wild-type (WT) macrophages were treated with or without inhibitors of PI3K (LY294002, 10uM) or MEK (U0126, 10 uM). Protein lysate of macrophages treated with thapsigargin (Thap, 5 uM) was used as positive controls. The levels of indicated proteins were determined by Western analysis. (Right) Primary macrophages isolated from Western-type diet-fed *ob/ob*;*Ldlr*<sup>-/-</sup> and *Ldlr*<sup>-/-</sup> mice receiving PBS or exendin-4 in vivo were used for the analysis of ATF-3 and P-PERK by Western analysis. **(F and G)** Atherosclerotic aortic sections of *ob/ob*;*Ldlr*<sup>-/-</sup> and *Ldlr*<sup>-/-</sup> mice treated with PBS or exendin-4 were probed with antibody against ATF-3 or P-PERK, or control antibody followed by appropriate Alexa fluor-conjugated secondary antibody. Representative images of ATF-3 or P-PERK (green) overlaid with Hoechst-stained nuclei (blue) are shown. **(H)** Double immunofluorescence staining using antibodies against UPR markers ATF-3 or active caspase 3 and macrophage marker Mac-3 in atherosclerotic lesions of *Insr*<sup>-/-</sup>.*Ldlr*<sup>-/-</sup> mice with in vivo treatment of PBS or exendin-4. The data of ATF-3 or caspase 3 positive macrophages are expressed as % of total macrophages in the same lesion areas. \*, P<0.05 for exendin-4 vs. saline-treated *Insr*<sup>-/-</sup>.*Ldlr*<sup>-/-</sup> mice. n=3.



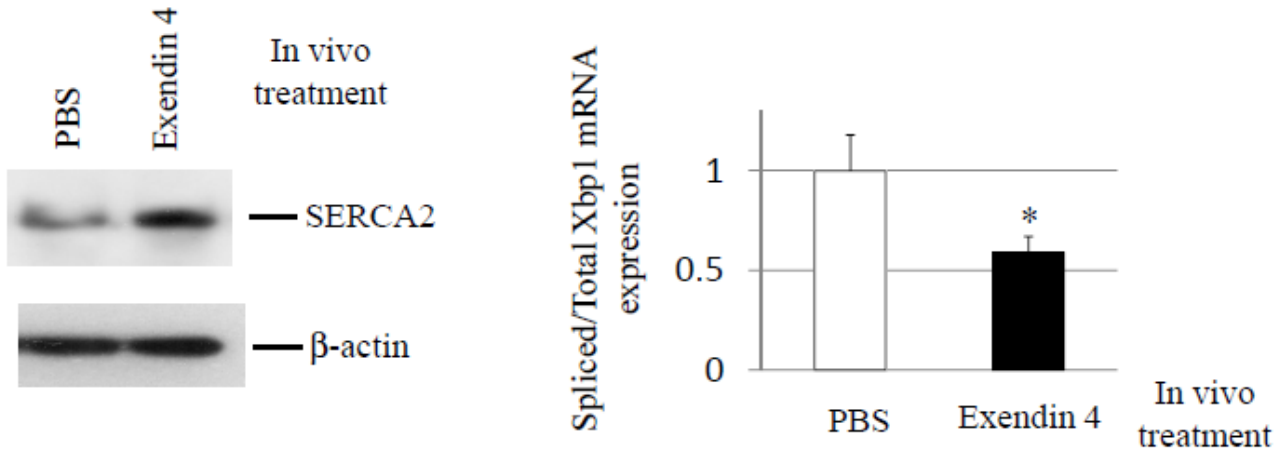
**B.**

| Treatment           | <i>ob/ob</i> . <i>Ldlr</i> <sup>-/-</sup> |            |
|---------------------|---|------------|
|                     | PBS                                       | Exendin 4  |
| TG (mg/dl)          | 1500 ± 229                                | 1295 ± 364 |
| Cholesterol (mg/dl) | 1738 ± 322                                | 1519 ± 104 |

SUPPLEMENTARY DATA

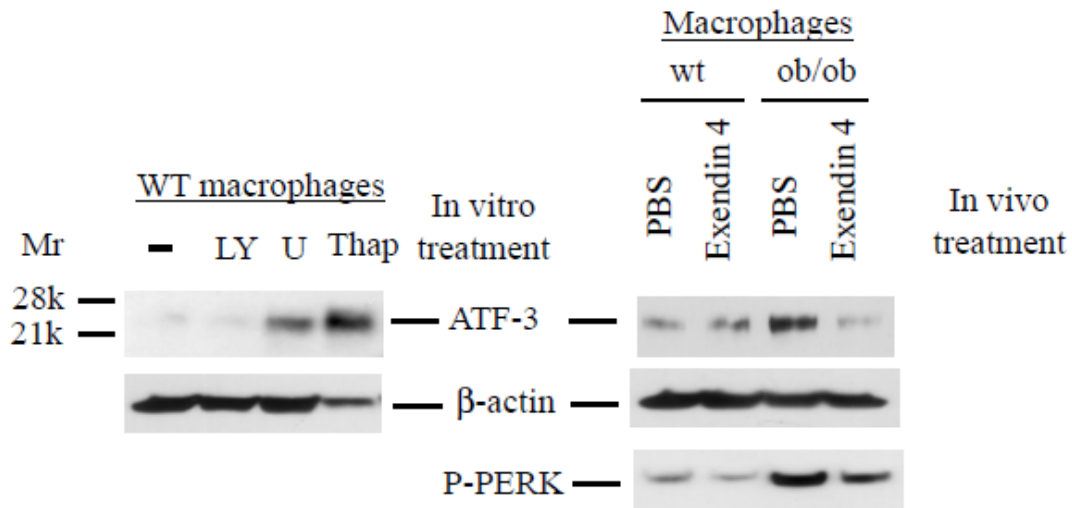
D.

ob/ob Peritoneal macrophages



E.

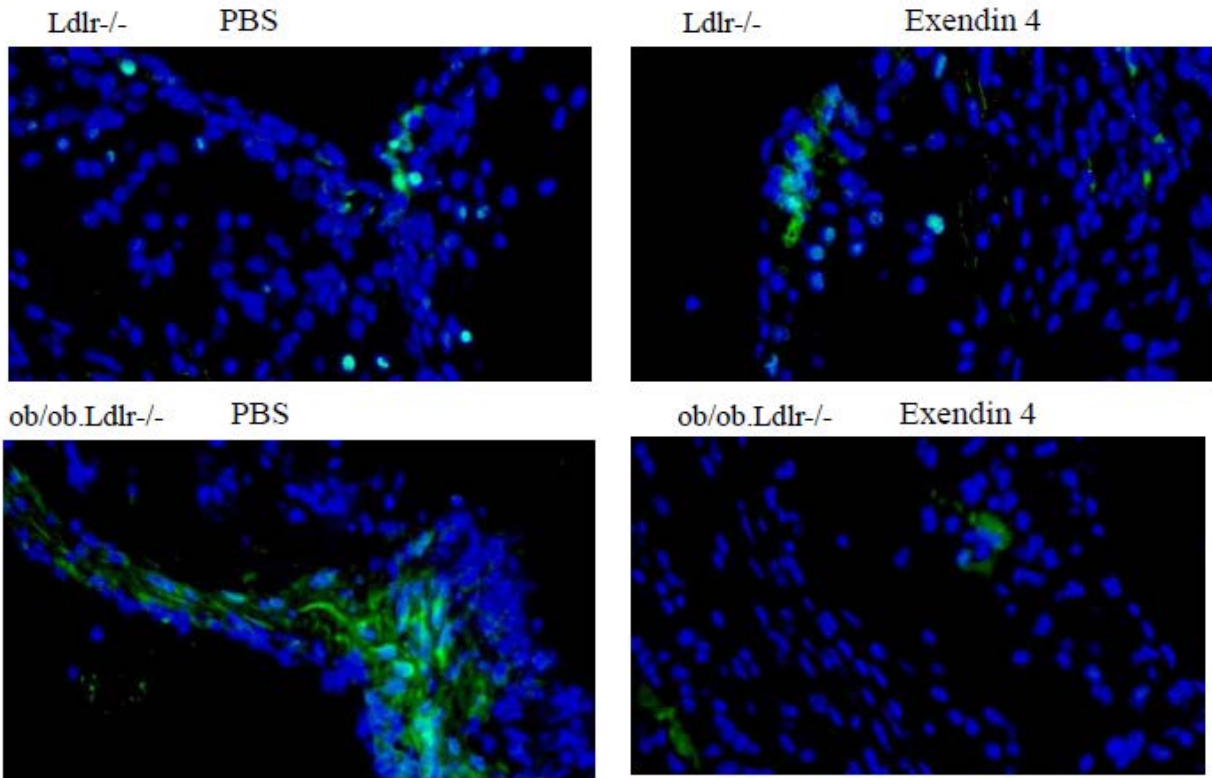
Peritoneal macrophages



SUPPLEMENTARY DATA

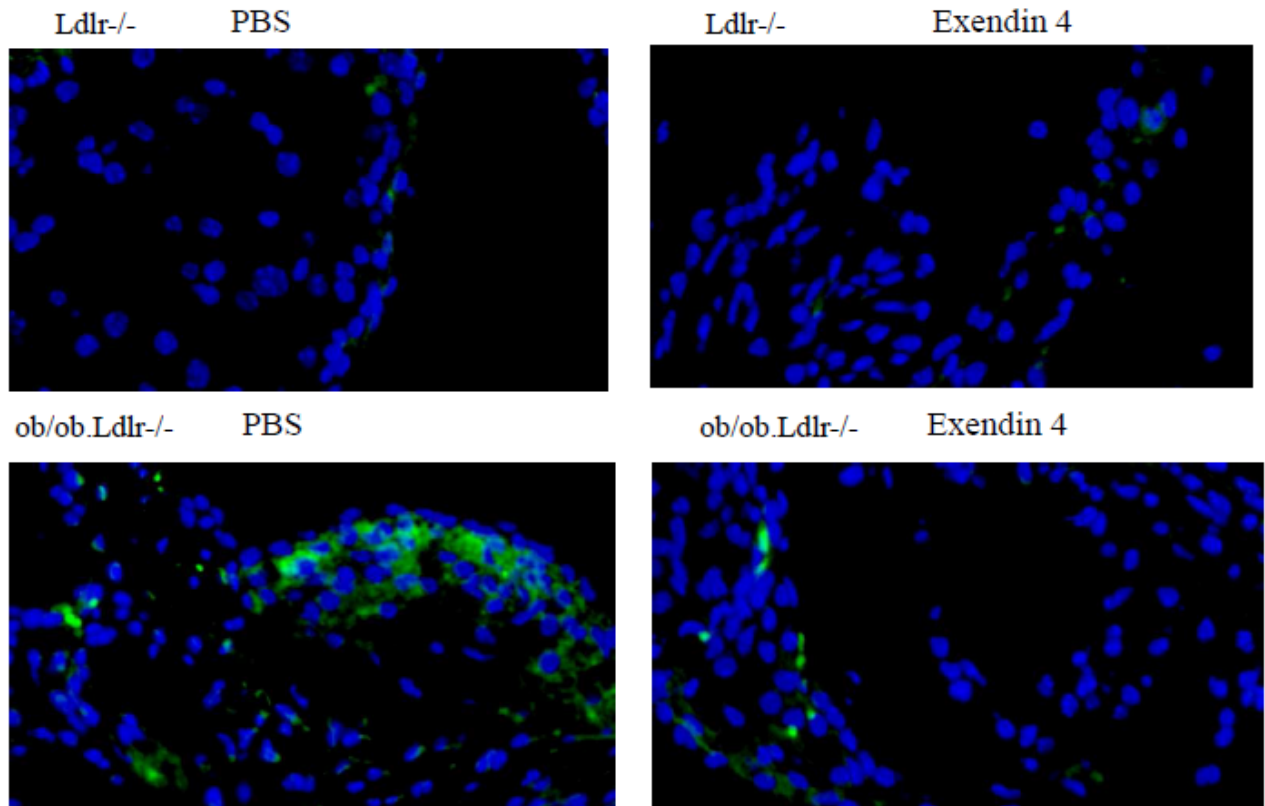
F.

ATF-3 Hoechst



G.

P-PERK Hoechst



H.

